

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

GIANNI=1

**TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371**

U.S. APPLICATION NO. (If known, see 37 CFR 1.52)

09/869612INTERNATIONAL APPLICATION NO.
PCT/EP99/10470INTERNATIONAL FILING DATE
30 December 1999PRIORITY CLAIMED
30 December 1998

TITLE OF INVENTION

HUMAN-GROWTH HORMONE TO STIMULATE MOBILIZATION OF PLURIPOTENT...

APPLICANT(S) FOR DO/EO/US

Alessandro Massimo GIANNI

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ The US has been elected in a Demand by the expiration of 19 months from the priority date (PCT Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☐ is attached hereto (required only if not transmitted by the International Bureau).
 - b. ☒ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made, however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern document(s) or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98
12. ☐ An Assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A **FIRST** preliminary amendment.
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
14. ☐ A substitute specification
15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information
 - ☒ Courtesy copy of the International Application as filed.
 - ☒ Courtesy copy of the first page of the International Publication (WO 00/40260)
 - ☒ **Courtesy copy of the International Preliminary Examination Report with annexes containing claims 1-55 to be substituted for original claims 1-60 for examination in this case.**
 - ☒ Formal drawings, 2 sheets, Figures 1-2
 - ☒ Courtesy Copy of the International Search Report.

☒ The application is (or will be) assigned to APPLIED RESEARCH SYSTEMS ARS HOLDING N.V. whose address is Pietermaai 15, Curacao (AN)

U.S. APPLICATION NO (If known, see 37 CFR 1.5)	International Application No	Attorney's Docket No
09/869612	PCT/EP99/10470	GIANNI=1

17. [xx] The following fees are submitted:

BASIC NATIONAL FEE (37 CFR 1.492 (a)(1) -(5):	
Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO.	\$1000.00
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO...	\$860.00
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO.. . . .	\$710.00
International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4).. . . .	\$690.00
International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4).	\$100.00

CALCULATIONS PTO USE ONLY	
ENTER APPROPRIATE BASIC FEE AMOUNT =	\$ 860.00
Surcharge of \$130.00 for furnishing the oath or declaration later than [] 20 [X] 30 months from the earliest claimed priority date (37 CFR 1.492(e)).	\$ 130.00
Claims as Originally Presented	
Total Claims	92 - 20 72 X \$18.00 \$1,296.00
Independent Claims	09 - 3 6 X \$80.00 \$ 480.00
Multiple Dependent Claims (if applicable)	+ \$270.00 \$ 270.00
TOTAL OF ABOVE CALCULATIONS =	\$3,036.00
Claims After Post Filing Prel Amend	
Total Claims	- 20 X \$18.00 \$
Independent Claims	- 3 X \$78.00 \$
TOTAL OF ABOVE CALCULATIONS =	\$3,036.00
Reduction of ½ for filing by small entity, if applicable. Applicant claims small entity status. See 37 CFR 1.27	\$
SUBTOTAL =	\$3,036.00
Processing fee of \$130.00 for furnishing the English translation later than [] 20 [] 30 months from the earliest claimed priority date (37 CFR 1.492(f))	\$
TOTAL NATIONAL FEE =	\$3,036.00
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +	\$
TOTAL FEES ENCLOSED =	\$3,036.00
	Amount to be: \$
	refunded
	charged \$

a. [] A check in the amount of \$ _____ to cover the above fees is enclosed.

b. [X] Credit Card Payment Form (PTO-2038), authorizing payment in the amount of \$ 3,036.00, is attached.

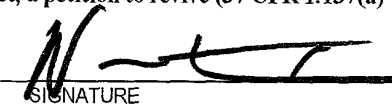
c. [] Please charge my Deposit Account No **02-4035** in the amount of \$ _____ to cover the above fees.
A duplicate copy of this sheet is enclosed

d. [XX] The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No **02-4035**. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO.

BROWDY AND NEIMARK, P.L.L.C.
624 NINTH STREET, N.W., SUITE 300
WASHINGTON, D.C. 20001
TEL: (202) 628-5197
FAX: (202) 737-3528
Date of this submission: **July 2, 2001**



SIGNATURE
Norman J. Latker
NAME
19,963
REGISTRATION NUMBER

INVENTOR INFORMATION

09/869612

Inventor One Given Name:: Alessandro M
Family Name:: GIANNI
Postal Address Line One:: Via Ludovico Muratori, 29
City:: Milano
Country:: Italy
Postal or Zip Code:: I-20135
Citizenship Country:: Italy

JC18 Rec'd PCT/PTO 02 JUL 2001

CORRESPONDENCE INFORMATION

Correspondence Customer Number:: 001444
Fax One:: 202-737-3528
Electronic Mail One:: Mail@BrowdyNeimark.com

APPLICATION INFORMATION

Title Line One:: HUMAN GROWTH HORMONE TO STIMULATE MOBILI
Title Line Two:: ZATION OF PLURIPOTENT HEMATOPOIETIC STEM
Title Line Three:: CELLS
Total Drawing Sheets:: 2
Formal Drawings?:: Yes
Docket Number:: GIANNI=1
Secrecy Order in Parent Appl.?:: No

REPRESENTATIVE INFORMATION

Representative Customer Number:: 1444

CONTINUITY INFORMATION

This application is a:: 371 OF
> Application One:: PCT/EP99/10470
Filing Date:: 12-30-1999

PRIOR FOREIGN APPLICATIONS

Foreign Application One:: 98124834.7
Filing Date:: 12-30-1998
Country:: Europe
Priority Claimed:: Yes

Source:: PrintEFS Version 1.0.1

1444 99 470 371 OF

JC18 Rec'd PCT/PTO 02 JUL 2001

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:)	Art Unit:
Alessandro Massimo GIANNI)	
)	
IA No.: PCT/EP99/10470)	
)	Washington, D.C.
IA Filed: 30 December 1999)	
)	
U.S. App. No.:)	
(Not Yet Assigned))	
)	July 2, 2001
National Filing Date:)	
(Not Yet Received))	
)	
For: HUMAN GROWTH HORMONE...)	Docket No.: GIANNI=1

PRELIMINARY AMENDMENT

Honorable Commissioner for Patents and Trademarks
Washington, D.C. 20231

Sir:

Contemporaneous with the filing of this case and prior to calculation of the filing fee, kindly amend as follows:

IN THE SPECIFICATION

After the title please insert the following paragraph:

REFERENCE TO RELATED APPLICATIONS

--The present application is the national stage under 35 U.S.C. 371 of international application PCT/EP99/10470, filed 30 December 1999 which designated the United States, and which international application was published under PCT Article 21(2) in the English language.--

09/069612

In re of: Alessandro Massimo GIANNI (GIANNI=1)

REMARKS

The above amendment to the specification is being made to insert reference to the PCT application of which the present case is a U.S. national stage

Favorable consideration and allowance are earnestly solicited.

Respectfully submitted,
BROWDY AND NEIMARK, P.L.L.C.
Attorneys for Applicant

By: 

Norman J. Latker

Registration No. 19,963

NJL:wrd

Telephone No.: (202) 628-5197

Facsimile No.: (202) 737-3528

1

Human growth hormone to stimulate mobilization of
pluripotent hematopoietic stem cells

Bone marrow transplantation (BMT) is a clinical procedure in which pluripotent hematopoietic cells obtained from bone marrow are transplanted to a patient. BMT is the treatment of choice in several hematological disorders, including malignancies, Severe Combined Immune Deficiencies (SCIDs), congenitally or genetically determined hematopoietic abnormalities, anemia, aplastic anemia, leukemia and osteopetrosis (Fischer et al., 1998). In the last ten years, the use of BMT grew from less than 5'000 to more than 40'000 annually (Waters et al., 1998).

Under steady state condition, the majority of hematopoietic stem and progenitor cells reside in the bone marrow and only a low number of these cells are detectable in peripheral blood. However, additional stem cells can be mobilized into the peripheral blood by treatment with myelosuppressive agents and/or certain hematopoietic growth factors (Van Hoef, 1998). Studies have demonstrated that peripheral blood stem cells (PBSC) infused in a host exhibits enhanced potential for engraftment as compared to bone marrow-derived stem and progenitor cells (Gianni et al., 1989 ; Larsson et al., 1998). Thus, PBSC mobilized by chemotherapy, hematopoietic growth factors or the combination of these modalities are currently used in both autologous and nonautologous transplantation settings (Van Hoef, 1998 ; Anderlini and Korbaling, 1997). In the case of nonautologous transplantation, the donors of stem cells are normal individuals and the procedure for mobilization of stem cells into the blood stream has to be achieved with minimal discomfort. In this case, stem cells

mobilization with hematopoietic growth factors is preferred to the treatment with antitlastic drugs (i.e. cyclophosphamid).

Several hematopoietic growth factors, such as G-CSF, EPO and CSF have been studied as mobilizing agents and are currently used to increase the number of PBSC prior to leukapheresis (Henry, 1997; Weaver and Testa, 1998). Treatments aimed at stimulating the overall hematopoiesis may be of great interest to mobilize a large set of progenitor cells and stem cells. Increased mobilization of stem cells is extremely valuable in the context of hematopoietic stem cells transplantation by reducing the number of leukapheresis required to collect sufficient amount of hematopoietic stem cells to be transplanted.

The first part of the invention provides a new mobilising agent used to increase the number of circulating cells capable of regenerating hematopoiesis in vivo in an individual.

The new mobilizing agent of the invention is growth hormone and especially Human Growth Hormone (hGH) or one of its derivatives or any factor inducing growth hormone release.

Unless it is otherwise specified, the term « GH » means Growth Hormone, one of its derivatives or any factor inducing growth hormone release within the context of the invention.

It has now been found that, by administering growth hormone and especially Human Growth Hormone (hGH) or one of its derivatives or any factor inducing growth hormone release, a mobilization of cells capable of regenerating hematopoiesis in vivo is obtained in the peripheral blood. Therefore, growth hormone and especially human Growth Hormone (hGH) or one of its derivatives or any factor inducing growth hormone release, administered

alone or in combination with other factors, represents a new method or use to mobilize cells capable of regenerating hematopoiesis in vivo to the peripheral blood.

Human Growth Hormone (hGH), also known as somatotropin is a protein hormone produced and secreted by the somatotropic cells of the anterior pituitary. hGH plays a key role in somatic growth through its effects on the metabolism of proteins, carbohydrates and lipids. In addition to its effects on somatic growth, hGH has been shown to stimulate blood cells in vitro (Derfalvi et al., 1998 ; Merchav et al; 1988), to increase erythrocytes and hemoglobin counts (Valerio et al. , 1997 ; Vihervuori et al. , 1996), to enhance both proliferation and Ig production in plasma cell lines (Kimata and Yoshida, 1994) and to stimulate CD8⁺ cell counts and, to a lesser extent CD4⁺ cell counts (Geffner, 1997).

The methods and uses of the invention which use the mobilising agent of the invention have several advantages :

- There is a low number of circulating cells capable of regenerating hematopoiesis. This number is considered insufficient to provide a cells engraftment dose by single or multiple apheresis in a reasonable time period. Methods and uses of the invention solve this problem by a temporary peripheralization of said cells and subsets into the circulating blood which is widely used to significantly increase in the blood the yield of circulating cells capable of regenerating hematopoiesis in vivo, thus minimizing the number of aphereses needed to achieve an engraftment dose.

- Other advantages of the methods and uses of the invention include the possibility of :

a) circumventing the need of general anesthesia,

- b) harvesting even if iliac bones are damaged by previous radiotherapy or infiltrated with malignant cells,
 - c) achieving restoration of sustained hematopoietic functions more rapidly than with BM derived progenitor cells.
 - d) achieving restoration of sustained hematopoietic functions more rapidly and effectively than without a pre-treatment including a method or a use of the invention.
- Generally, methods and uses of the invention are effective and safe to mobilize to peripheral blood cells capable of regenerating hematopoiesis in vivo.
 - Methods and uses of the invention are not toxic in view of main parameters of toxicity which are for example tumor growth, clinical and instrumental symptoms, or laboratory tests for cardiac, liver and renal function.
 - The increased mobilization of circulating cells capable of regenerating hematopoiesis in vivo obtained with the methods and uses of the invention is extremely valuable in the context of hematopoietic stem cells transplantation by reducing the number of leukapheresis required to collect sufficient amount of hematopoietic cells to be transplanted.
 - Methods and uses of the invention lead to a reduction of the volume of blood required to be processed during the apheresis or leukapheresis procedure in order to obtain the specified target number of cells. The advantages of processing a reduced volume of blood are that the patient spends less time on the cell separating machine, that it reduces the toxicity of the procedure, particularly in terms of the volume of anticoagulant to which the patient would be exposed during the procedure, that it reduces the machine and the operator's time.
 - Furthermore, the transplantation of a population of blood cells enriched with cells capable of regenerating

hematopoiesis in vivo, which population is obtained from the peripheral blood by the methods or uses of the invention has the effect to enhance reconstitution of recipient's hematopoietic and immune systems following myeloablative or antitlastic therapies.

In a first aspect, the invention concerns a method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising :

- a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo,
- b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

The method of the invention thus produces a population of cells capable of regenerating hematopoiesis in vivo, this population being destined for transplantation in the same or in different individuals.

Thus, the invention concerns a method of preparation of a population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo comprising :

- a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo,
- b) isolating a population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

In a further embodiment, the invention relates to a method of isolating an increased number of circulating cells capable of regenerating hematopoiesis in vivo from a donor comprising :

- a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release alone, or in combination with other hematopoietic growth factors, to the subject in an amount sufficient to induce mobilization of cells capable of regenerating hematopoiesis in vivo to the peripheral blood,
- b) isolating a population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

In another embodiment, the invention concerns a method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising :

- a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing the growth hormone release in an amount sufficient to induce in said donor the mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo,
- b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor or isolating a population of blood cells enriched with circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

Step b) [i.e. « isolating a population of (blood cells enriched with) circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor »] of the methods or uses of the invention may correspond to the operation of removing

peripheral blood from the donor wherein the number of cells capable of regenerating hematopoiesis in vivo has been increased by administration of growth hormone or one of its derivatives or any factor inducing growth hormone alone or in combination with other factors.

An amount sufficient to increase the number of circulating cells capable of regenerating hematopoiesis in vivo, an amount sufficient to induce the mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo or an amount sufficient to induce mobilization of cells capable of regenerating hematopoiesis in vivo to the peripheral blood can be administered in one or several doses during one or several days.

The operation of removing peripheral blood from the donor may correspond to leukapheresis. Leukapheresis is a procedure, in which, leukocytes are removed from the withdrawn blood and the remainder of the blood is retransfused into the donor.

Cells capable of regenerating hematopoiesis in vivo present in the isolated population of blood cells can be further purified in order to increase the concentration of said cells. Said purification may be done by positive selection of CD34 positive cells.

In a further embodiment, the invention concerns a method of preparation of a donor of circulating cells, which cells are capable of regenerating hematopoiesis in vivo comprising the administration to said donor of a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase the number of circulating cells capable of regenerating hematopoiesis in vivo in said donor.

In another embodiment, the invention concerns a method for increasing the number of circulating cells

capable of regenerating hematopoiesis in vivo in a donor by administration to said donor of a composition comprising growth hormone or one of its derivatives or any factor inducing the growth hormone release.

The term "increased" or "increase" and the term "enriched" generally mean in the context of the invention that the "increased" or "enriched" parameter (number) has a value which is above the standard value of this parameter. The standard value of the parameter is measured in a body or in a sample of a body which has not received any mobilising agent of cells capable of regenerating hematopoiesis in vivo. The standard value of the number of CD34⁺ cells per microliter of blood is for example 3.8 (+ or - 3.2) cells per microliter of peripheral blood (Anderlini et al., 1997).

The circulating cells capable of regenerating hematopoiesis in vivo may be CD34⁺ cells.

The frequency of CD34⁺ cells in the blood may be measured by FACScan measurements (Siena et al., 1989 & 1991).

The increased number of CD34⁺ cells in the peripheral blood of the donor or the level of enrichment of CD34⁺ cells in the isolated preparation of blood cells may be more than 10, 25, 34 or 80 CD34⁺ cells per microliter of peripheral blood.

The increased number of CD34⁺ cells in the peripheral blood of the donor or the level of enrichment of CD34⁺ cells in the isolated preparation of blood cells may be at least 2×10^6 CD34⁺ cells per kilogram of recipient body weight, or at least 4×10^6 CD34⁺ cells per kilogram of recipient body weight or at least 8×10^6 CD34⁺ cells per kilogram of recipient body weight.

The increased number of CD34⁺ cells in the peripheral blood of the donor or the level of enrichment of CD34⁺ cells in the isolated preparation of blood cells

may be at least 2×10^6 , 4×10^6 , 5×10^6 , 6×10^6 , 8×10^6 or 15×10^6 CD34⁺ cells per kilogram of donor body weight.

There is a correlation between the number of CD34⁺ cells required for transplantation and the corresponding GM-CFC activity which can be measured (Weaver et al., 1998). Therefore, the increased number of circulating cells capable of regenerating hematopoiesis in vivo or the level of enrichment of cells capable of regenerating hematopoiesis in vivo in the isolated preparation of blood cells may correspond to at least 1×10^5 GM-CFC per kilogram of donor or recipient body weight.

The number of CD34⁺ cells in the blood correlates well with CFU-GM (Siena et al., 1991). CFU-GM is the colony forming unit, granulocyte and macrophage. Therefore, the increased number of circulating cells capable of regenerating hematopoiesis in vivo or the level of enrichment of cells capable of regenerating hematopoiesis in vivo in the isolated preparation of blood cells may correspond to at least 500 CFU-GM per milliliter of peripheral blood.

With the same reasoning, the increased number of circulating cells capable of regenerating hematopoiesis in vivo or the level of enrichment of cells capable of regenerating hematopoiesis in vivo in the isolated preparation of blood cells may correspond to an increased level of CFU-C, CFU-Meg or BFU-E. CFU-C is the colony forming unit, culture ; CFU-Meg is the colony forming unit, megakaryocyte and BFU-E is the burst forming unit, erythroid.

The number of CD34⁺ cells in the blood correlates well with the white blood cell count. Therefore, the increased number of circulating cells capable of regenerating hematopoiesis in vivo or the level of enrichment of cells capable of regenerating hematopoiesis in vivo in the isolated preparation of blood cells may

correspond to at least 1000 white blood cells per microliter of peripheral blood.

The circulating cells capable of regenerating hematopoiesis in vivo may be CD34⁺/CD33⁺ cells and/or CD34⁺/CD38⁺ cells and/or CD34⁺/Thy-I cells and/or CD34⁺/Thy-I/CD38⁺ cells and/or CD33⁺ cells and/or bone-marrow stem cells and/or progenitor cells and/or long-term culture initiating cells (LTC-IC) and/or cells that fulfill self renewal potential and/or cells that fulfill pluripotential characteristics and/or cells that initiate long term bone marrow culture and/or cells that can generate multiple cell lineages. Cell lineages may be fully differentiated blood cells.

The CD34⁺/CD38⁺ cells and CD34⁺/Thy-I cells and CD34⁺/Thy-I/CD38⁺ cells are recited for example in Anderlini et al (see references). The CD34⁺/CD33⁺ cells and the CD33⁺ cells are recited for example in Siena et al ; 1991 (see references). The long-term culture initiating cells (LTC-IC) are recited for example in Heather et al (see references). Cells that fulfill self renewal potential and/or cells that fulfill pluripotential characteristics and/or cells that initiate long term bone marrow culture are recited for example in Anderlini et al (see references).

In a further embodiment, the invention relates to the following uses :

- Use of growth hormone, one of its derivatives, or any factor inducing growth hormone release for increasing or expanding the number of circulating cells capable of regenerating hematopoiesis in vivo.
- Use of growth hormone, one of its derivatives or any factor inducing growth hormone release for peripheralizing cells capable of regenerating hematopoiesis in vivo.

- Use of growth hormone, one of its derivatives, or any factor inducing growth hormone release to prepare a medicament or a composition for increasing or expanding the number of circulating cells capable of regenerating hematopoiesis in vivo.
- Use of growth hormone, one of its derivatives or any factor inducing growth hormone release to prepare a medicament or a composition for peripheralizing cells capable of regenerating hematopoiesis in vivo.
- Use according to any one of the preceding uses wherein the circulating cells capable of regenerating hematopoiesis in vivo are CD34⁺ cells.
- Use according to the preceding use wherein the increased number of CD34⁺ cells is more than 10, 25, 34 or 80 CD34⁺ cells per microliter of peripheral blood.
- Use according to any one of the preceding uses wherein the increased number of CD34⁺ cells is at least 2×10^6 CD34⁺ cells per kilogram of recipient body weight, or at least 4×10^6 CD34⁺ cells per kilogram of recipient body weight or at least 8×10^6 CD34⁺ cells per kilogram of recipient body weight.
- Use according to any one of the preceding uses wherein the increased number of CD34⁺ cells is at least 2×10^6 , 4×10^6 , 5×10^6 , 6×10^6 , 8×10^6 or 15×10^6 CD34⁺ cells per kilogram of donor body weight.
- Use according to any one of the preceding uses wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to at least 1×10^5 GM-CFC per kilogram of donor or recipient body weight.
- Use according to any one of the preceding uses wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to at least 500 CFU-GM per milliliter of peripheral blood.

- Use according to any one of the preceding uses wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to an increased level of CFU-C, CFU-Meg or BFU-E.
- Use according to any one of the preceding uses wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo substantially corresponds to a white blood cell count which is at least 1000 cells per microliter of peripheral blood.
- Use according to any one of the preceding uses wherein the circulating cells capable of regenerating hematopoiesis in vivo are CD34⁺/CD33⁺ cells and/or CD34⁺/CD38⁺ cells and/or CD34⁺/Thy-I cells and/or CD34⁺/Thy-I/CD38⁺ cells and/or CD33⁺ cells and/or stem cells and/or progenitor cells and/or long-term culture initiating cells (LTC-IC) and/or cells that fulfill self renewal potential and/or cells that fulfill pluripotential characteristics and/or cells that initiate long term bone marrow culture.
- Use according to any one of the preceding uses wherein the medicament or composition comprises further one or several compound(s) chosen among the following groups of compounds : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.
- Use according to any one of the preceding uses wherein the cytokines group comprises IL-1, IL-3, G-CSF, GM-CSF or SCF ; the chemokines group comprises MIP-1 α or thrombopoietin (TPO) ; the monoclonal antibodies group comprises anti-VLA-4 antibodies.
- Use according to any one of the preceding uses wherein the medicament or composition comprises Growth Hormone and G-CSF.
- Use according to any one of the preceding uses wherein GH and G-CSF are administered separately and/or simultaneously.

- Use according to any one of the preceding uses wherein Growth Hormone is administered in an amount of around $33\mu\text{g}$ per kilogram of body weight.
- Use according to any one of the two preceding uses wherein the G-CSF is administered in an amount of around $5\mu\text{g}$ or around $10\mu\text{g}$ per kilogram.
- Use according to any one of the two preceding uses wherein the administration is made by intravenous or subcutaneous route.
- Use according to any one of the preceding uses wherein the administration is made by parenteral, subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal or buccal routes.
- Use according to any one of the preceding uses wherein the administration is daily or three times a day.
- Use according to any one of the preceding uses wherein the administration of growth hormone is done three times a day and the administration of G-CSF is done daily.
- Use according to any one of the preceding uses wherein the administration is made over a period of 5 days or over a period of 10 days, until leukapheresis or until full recovery.
- Use according to any one of the preceding uses wherein the administration is made until leukapheresis or until full recovery.
- Use according to any one of the preceding uses wherein growth hormone is recombinant growth hormone.
- Use according to any one of the preceding uses wherein growth hormone is human growth hormone.

In a second aspect, the invention concerns a method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising :

- a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo,
- b) processing or isolating said reduced volume of blood ; and optionally
- c) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from said isolated volume.

Step b) or c) [isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from said isolated volume] of the methods or uses of the invention may correspond to the operation of removing peripheral blood from the donor wherein the number of cells capable of regenerating hematopoiesis in vivo has been increased by administration of growth hormone or one of its derivatives or any factor inducing growth hormone alone or in combination with other factors.

An amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo can be administered in one or several doses during one or several days.

The operation of removing peripheral blood from the donor may correspond to leukapheresis. Leukapheresis is a procedure, in which, leukocytes are removed from the withdrawn blood and the remainder of the blood is retransfused into the donor.

Cells capable of regenerating hematopoiesis in vivo present in the isolated population of blood cells can be further purified in order to increase the concentration

of said cells. Said purification may be done by positive selection of CD34 positive cells.

In another embodiment, the invention concerns a method of preparation of a donor of circulating cells, which cells are capable of regenerating hematopoiesis in vivo comprising the administration to said donor of a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo and/or to reduce the number of leukapheresis required to collect sufficient amount of circulating cells capable of regenerating hematopoiesis in vivo to be transplanted.

An amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo and/or to reduce the number of leukapheresis required to collect sufficient amount of circulating cells capable of regenerating hematopoiesis in vivo to be transplanted can be administered in one or several doses during one or several days.

The volume of blood required to be processed may be the volume of blood required to be processed during the apheresis or leukapheresis procedure.

In a further embodiment, the invention concerns a method for reducing the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo in a donor and/or for reducing the number of leukapheresis required to collect sufficient amount of circulating cells capable of regenerating hematopoiesis in vivo to be transplanted by administration

of a composition comprising the growth hormone or one of its derivatives or any factor inducing the growth hormone release to said donor .

The term "reduced" generally means in view of the invention that the "reduced" parameter (volume) has a value which is inferior to the standard value of this parameter.

The specified target number of circulating cells capable of regenerating hematopoiesis in vivo is at least 2×10^4 LTC-IC per kg of donor or recipient body, around or more than 2×10^6 CD34⁺ cells per kilogram of donor or recipient body weight, around or more than 4×10^6 CD34⁺ cells per kilogram of donor or recipient body weight or around or more than 8×10^6 CD34⁺ cells per kilogram of donor or recipient body weight.

The required volume of blood may be comprised in a range of about 30 to about 900 milliliters.

In another embodiment, the invention relates to the following uses :

- Use of growth hormone, one of its derivatives or any factor inducing growth hormone release for reducing the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo.
- Use of growth hormone, one of its derivatives or any factor inducing growth hormone release to prepare a medicament or a composition for reducing the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo.
- Use according to the preceding use wherein the specified target number of circulating cells capable of regenerating hematopoiesis in vivo is around or more than 2×10^4 LTC-IC per kg of donor or recipient body, around or more than 2×10^6 CD34⁺ cells per

kilogram of donor or recipient body weight, around or more than 4×10^6 CD34⁺ cells per kilogram of donor or recipient body weight or around or more than 8×10^6 CD34⁺ cells per kilogram of donor or recipient body weight.

- Use according to any one of the two preceding uses wherein the required volume of blood is comprised in a range of about 30 to about 900 milliliters.
- Use according to any one of the preceding uses wherein the medicament or composition comprises further one or several compound chosen among the following groups of compounds : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.
- Use according to any one of the preceding uses wherein the cytokines group comprises IL-1, IL-3, G-CSF, GM-CSF or SCF ; the chemokines group comprises MIP-1 α or thrombopoietin (TPO) ; the monoclonal antibodies group comprises anti-VLA-4 antibodies.
- Use according to any one of the preceding uses wherein the medicament or composition comprises Growth Hormone and G-CSF.
- Use according to any one of the preceding claims wherein GH and G-CSF are administered separately and/or simultaneously.
- Use according to any one of the preceding uses wherein Growth Hormone is administered in an amount of around 33 μ g per kilogram of body weight.
- Use according to any one of the two preceding uses wherein the G-CSF is administered in an amount of around 5 μ g or around 10 μ g per kilogram.
- Use according to any one of the two preceding uses wherein the administration is made by intravenous or subcutaneous route.
- Use according to any one of the preceding uses wherein the administration is made by parenteral,

subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal or buccal routes.

- Use according to any one of the preceding uses wherein the administration is daily or three times a day.
- Use according to any one of the preceding claims wherein the administration of growth hormone is done three times a day and the administration of G-CSF is done daily.
- Use according to any one of the preceding uses wherein the administration is made over a period of 5 days or over a period of 10 days, until leukapheresis or until full recovery.
- Use according to any one of the preceding claims wherein the administration is made until leukapheresis or until full recovery.
- Use according to any one of the preceding uses wherein the administration(s) is(are) made after chemotherapy, radiotherapy, myelosuppressive therapy, transplantation of cells capable of regenerating hematopoiesis in vivo or transplantation of bone-marrow.
- Use according to any one of the preceding uses wherein the administration(s) begin(s) around 7 days after the beginning of the chemotherapeutic treatment or around 2 days after the end of the chemotherapeutic treatment.
- Use according to any one of the preceding uses wherein the growth hormone is recombinant growth hormone.
- Use according to any one of the preceding uses wherein the growth hormone is human growth hormone.

In this application :

- The term "circulating" may be replaced by the term "blood" or "peripheral blood".

- The term "preparation" in the expression "method of preparation" may be replaced by "pre-treatment" or by "preparation for blood extraction or leukapheresis".
- A "donor" as recited in the methods or uses of the invention may be a human or an animal, a healthy or a sick individual (patient). Said animal is preferably a mammal and may be chosen from domestic animals such as dogs, cats etc. or animals such as horses, cattle, sheep.
- The term « hematopoiesis » can mean the formation of the blood cells.
- The term "Growth hormone" encompasses human growth hormone (hGH) and all the homologous proteins of human growth hormone of different species and all the homologs of human growth hormone in species other than human. Species other than human may be any sort of domestic animal or horse for example.

In a preferred embodiment, growth hormone is human growth hormone. Human growth hormone (hGH), also known as somatotropin is a protein hormone produced and secreted by the somatotropic cells of the anterior pituitary. hGH plays a key role in somatic growth through its effects on the metabolism of proteins, carbohydrates and lipids.

Human growth hormone is a single polypeptide chain of 101 amino acids having two disulfide bonds, one between Cys-53 and Cys-165, forming a large loop in the molecule, and the other between Cys-182 and Cys-189, forming a small loop near the C-terminus.

The term « derivative » in the expression « derivatives of growth hormone » signifies in the context of the invention, molecules which differ structurally from GH but which conserve the function of GH with respect to its direct or indirect effect on the metabolism of proteins, carbohydrates and lipids and/or its mobilisation effect and/or recovery effect (i.e.

« mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo »)

Derivatives of human growth hormone (hGH) included in the invention include naturally-occurring derivatives, variants and metabolic products, degradation products primarily of biosynthetic hGH and engineered derivatives of hGH produced by genetic methods. Any derivative of hGH can be used for the purpose of the present invention as long as it retains the biological activity of hGH in view of the invention.

Examples of derivatives are splice variants, oligomers, aggregates, proteolytic cleavage products, variants having substitutions, insertions or deletions of one or more amino acids etc.

Methionyl hGH is an example of derivative of hGH which is produced through recombinant DNA technology. This compound is actually a derivative of hGH having one additional methionine residue at its N-terminus (Goeddel et al., 1979).

Another example of derivative of hGH is a naturally occurring variant of hGH called 20-K-hGH which has been reported to occur in the pituitary as well as in the bloodstream (Lewis et al, 1978 ; Lewis et al, 1980). This compound, which lacks the 15 amino acid residues from Glu-32 to Gln-46, arises from an alternative splicing of the messenger ribonucleic acid (DeNoto et al., 1981).

Another example of derivative of hGH is acetylated at the N-terminus (Lewis et al., 1979).

Human growth hormone may further be in a monomeric, dimeric and higher molecular weight oligomeric form or in a mixture of said forms.

Human growth hormone may be in aggregated forms found both in the pituitary and in the circulation (Stolar et al., 1984 ; Stolar and Baumann, 1986).

The dimeric form of hGH may be of distinct types :

- a disulfide dimer connected through interchain disulfide bonds (Lewis et al., 1977),
- a covalent or irreversible dimer that is detected on sodium dodecylsulfate-polyacrylamide gels and that is not a disulfide dimer (Bewley and Li, 1975), and
- a non-covalent dimer which is easily dissociated into monomeric hGH by treatment with agents that disrupt hydrophobic interactions in proteins (Becker et al., 1987),
- a dimeric complex with Zn^{2+} (Cunningham et al., 1991).

Scatchard analysis has revealed that two Zn^{2+} ions associate per hGH dimer in a cooperative fashion, and this Zn^{2+} -hGH dimeric complex was found to be more stable to denaturation than monomeric hGH (Cunningham et al., 1991).

A number of derivatives of hGH arise from proteolytic modifications of the molecule. The primary pathway for the metabolism of hGH involves proteolysis. The region of hGH around residues 130-150 is extremely susceptible to proteolysis, and several derivatives of hGH having nicks or deletions in this region have been described (Thorlacius-Ussing, 1987). This region is in the large loop of hGH, and cleavage of a peptide bond there results in the generation of two chains that are connected through the disulfide bond at Cys-53 and Cys-

165. Many of these two-chain forms are reported to have increased biological activity (Singh et al., 1974).

Many derivatives of human growth hormone have been generated artificially through the use of enzymes. The enzymes trypsin and subtilisin, as well as others, have been used to modify hGH at various points throughout the molecule (Lewis et al., 1977). One such derivative, called two-chain anabolic protein (2-CAP), was formed through the controlled proteolysis of hGH using trypsin.

Another example of derivative of hGH is deamidated hGH. Asparagine and glutamine residues in proteins are susceptible to deamidation reactions under appropriate conditions. An example of deamidated hGH is pituitary hGH which has been shown to undergo this type of reaction, resulting in conversion of Asn-152 to aspartic acid and also, to a lesser extent, conversion of Gln-137 to glutamic acid (Lewis et al., 1981). Another example of deamidated hGH is Biosynthetic hGH which is known to degrade under certain storage conditions, resulting in deamidation at a different asparagine (Asn-149). This is the primary site of deamidation, but deamidation at Asn-152 is also seen (Becker et al., 1988). Deamidation at Gln-137 has not been reported in biosynthetic hGH.

Another example of derivative of hGH is sulfoxide hGH. Methionine residues in proteins are susceptible to oxidation, primarily to the sulfoxide. Both pituitary-derived and biosynthetic hGH undergo sulfoxidations at Met-14 and Met-125 (Becker et al., 1988). Oxidation at Met-170 has also been reported in pituitary but not biosynthetic hGH.

Another example of derivative of hGH is truncated forms of hGH which have been produced, either through the actions of enzymes or by genetic methods. 2-CAP, generated by the controlled actions of trypsin, has the first eight residues at the N-terminus of hGH removed.

Other truncated versions of hGH have been produced by modifying the gene prior to expression in a suitable host. The first 13 residues have been removed to yield a derivative having distinctive biological properties in which the polypeptide chain is not cleaved (Gertler et al., 1986).

hGH and its derivatives may be produced by recombinant DNA technology which permits production of an unlimited supply of hGH in a number of different systems. Purification of hGH or its derivatives from the culture medium is facilitated by low amounts of contaminating proteins present. In fact, it has been shown that hGH can be purified on a laboratory scale by a single purification step on a reversed-phase HPLC column.

Recombinant hGH is generally marketed as vials containing hGH plus additional excipients, e.g., glycine and mannitol, in a lyophilized form. A companion diluent vial is provided, allowing the patient to reconstitute the product to the desired concentration prior to administration of the dose.

In general, no significant differences have been observed in the pharmacokinetics or biological activities of recombinant natural sequence hGH, recombinant N-methionyl-hGH, or pituitary-derived material in humans (Moore et al., 1988 ; Jorgensen et al. , 1988).

The human growth hormone as used in the present invention can include functional derivatives as noted above, as well as other types of derivatives, fragments, variants, analogs, or chemical derivatives. A functional derivative retains at least a portion of the amino acid sequence of hGH which permits its utility in accordance with the present invention, namely mobilization of circulating cells capable of regenerating hematopoiesis in vivo for example.

In the meaning of the invention, a « derivative » may be :

- A "fragment" of the human growth hormone according to the present invention refers to any subset of the molecule, that is, a shorter peptide.
- A "variant" of the human growth hormone according to the present invention refers to a molecule which is substantially similar to either the entire peptide or a fragment thereof. Variant peptides may be conveniently prepared by direct chemical synthesis of the variant peptide, using methods well known in the art.

Alternatively, amino acid variants of hGH can be prepared by mutations in the cDNA encoding the synthesized hGH derivatives. Such variants comprise deletions, insertions or substitution of residues within the amino acid sequence. Any combination of deletions, insertions, and substitutions may also be made, provided that the final construct possesses the desired activity.

At the genetic level, these variants ordinarily are prepared by site-directed mutagenesis (as exemplified by (Adelman et al., 1983)) of nucleotides in the DNA encoding the peptide molecule, thereby producing DNA encoding the variant, and thereafter expressing the DNA in recombinant cell culture. The variants typically exhibit the same biological activity as the non-variant peptide.

- An "analog" of human growth hormone according to the present invention refers to a non-natural molecule which is substantially similar to either the entire molecule or to an active fragment thereof.
- A "chemical derivative" of human growth hormone according to the present invention contains additional chemical moieties not normally part of the human growth hormone derivative amino acid sequence. Covalent modifications of the amino acid sequence are included

within the scope of this invention. Such modifications may be introduced into the human growth hormone by reacting targeted amino acid residues of the peptide with an organic derivatizing agent that is capable of reacting with selected side chains or terminal residues.

The types of substitutions which may be made in the human growth hormone according to the present invention may be based on analysis of the frequencies of amino acid changes between a homologous protein of different species. Based upon such analysis, conservative substitutions may be defined herein as exchanges within one of the following five groups :

- I : Small, aliphatic, nonpolar or slightly polar residues : Ala, Ser, Thr, Pro, Gly
- II : Polar, negatively-charged residues and their amides : Asp, Asn, Glu, Gln
- III : Polar, positively-charged residues : His, Arg, Lys
- IV : Large, aliphatic non-polar residues : Met, Leu, Ile, Val, Cys
- V : Large aromatic residues : Phe, Try, Trp

Within the foregoing groups, the following substitutions are considered to be "highly conservative" :

- Asp/Glu
- His/Arg/Lys
- Phe/Tyr/Trp
- Met/Leu/Ile/Val

Semi-conservative substitutions are defined to be exchanges between two of groups (I)-(IV) above which are limited to supergroup (A), comprising (I), (II), and (III) above, or to supergroup (B), comprising (IV) and (V) above. Substitutions are not limited to the genetically encoded or even the naturally- occurring amino acids. When the epitope is prepared by peptide synthesis, the desired amino acid may be used directly.

Alternatively, a genetically encoded amino acid may be modified by reacting it with an organic derivatizing agent that is capable of reacting with selected side chains or terminal residues.

Cysteiny l residues most commonly are reacted with alpha- haloacetates (and corresponding amines), such as chloroacetic acid or chloroacetamide, to give carboxylmethyl or carboxyamidomethyl derivatives. Cysteiny l residues also are derivatized by reaction with bromotrifluoroacetone, alpha-bromo-beta-(5-imidazolyl)propionic acid, chloroacetyl phosphate, N-alkylmaleimides, 3-nitro-2-pyridyl disulfide, methyl-2-pyridyl disulfide, p-chloromercuribenzoate, 2-chloromercuri-4-nitrophenol, or chloro-7-nitrobenzo-2-oxa-1,3-diazole.

Histidyl residues are derivatized by reaction with diethylprocarbonate at pH 5.5-7.0 because this agent is relatively specific for the histidyl side chain. Parabromophenacyl bromide is also useful ; the reaction is preferably performed in 0.1 M sodium cacodylate at pH 6.0.

Lysiny l and amino terminal residues are reacted with succinic or other carboxylic acid anhydrides. Derivatization with these agents has the effect of reversing the charge of the lysiny l residues. Other suitable reagents for derivatizing alpha-amino acid-containing residues include imidoesters such as methyl picolinimide ; pyridoxal phosphate ; pyridoxal ; chloroborohydride; trinitrobenzenesulfonic acid ; O-methylisourea ; 2,4-pentanedione ; and transaminase-catalyzed reaction with glyoxylate.

Arginy l residues are modified by reaction with one or several conventional reagents, among them phenylglyoxal ; 2,3- butanedione ; and ninhydrin. Derivatization of arginine residues requires that the

reaction be performed in alkaline conditions because of the high pKa of the guanidine functional group. Furthermore, these reagents may react with the groups of lysine, as well as the arginine epsilon-amino group.

The specific modification of tyrosyl residues per se has been studied extensively, with particular interest in introducing spectral labels into tyrosyl residues by reaction with aromatic diazonium compounds or tetranitromethane. Most commonly, N-acetylimidazole and tetranitromethane are used to form O-acetyl tyrosyl species and e-nitro derivatives, respectively.

Carboxyl side groups (aspartyl or glutamyl) are selectively modified by reaction with carbodiimides (R'N-C-N-R') such as 1-cyclohexyl-3-[2-morpholinyl-(4-ethyl)]carbodiimide or 1-ethyl-3-(4-azonia-4,4-dimethylpentyl)carbodiimide. Furthermore, aspartyl and glutamyl residues are converted to asparaginyl and glutaminyl residues by reaction with ammonium ions.

Glutaminyl and asparaginyl residues are frequently deamidated to the corresponding glutamyl and aspartyl residues. Alternatively, these residues are deamidated under mildly acidic conditions. Either form of these residues falls within the scope of this invention.

While the present invention may be carried out with recombinant human growth hormone derivatives made by recombinant DNA technology, for instance in procaryotic or eucaryotic cells, these derivatives can also be made by conventional protein synthesis methods which are well known to those skilled in the art.

Growth hormone may be a protein, a peptide, a DNA molecule, a RNA molecule. DNA molecule and RNA molecule may encode hGH and all its derivatives including those recited above.

Growth hormone may preferably be recombinant growth hormone.

Determination of amounts of growth hormone, of one of its derivatives or of any factor inducing growth hormone release to be administered in a method or use of the invention described above is within the skill of the art.

Typical dosage of growth hormone, of one of its derivatives or of any factor inducing growth hormone release will start at about 1 microgram per kilogram of the patient weight per day and dose will be escalated until the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo) is reached.

The dosage of growth hormone, of one of its derivatives or of any factor inducing growth hormone release administered depends upon the age, sex, health and weight of the donor, type of previous or concurrent treatment, if any, frequency of the treatment and the nature of the effect desired.

Growth hormone or one of its derivatives may advantageously be administered in an amount comprised between 20 and 50 μg per kilogram of body weight, more particularly between 30 and 40 μg per kilogram of body weight.

A preferred dosage of Growth hormone or one of its derivatives to be administered is around 33 μg per kilogram of body weight.

Growth hormone or its derivatives or any factor inducing growth hormone release may be administered alone or in conjunction or association with other factors.

Growth hormone or its derivatives or any factor inducing growth hormone release may advantageously be present in a composition which comprises further one or several compound(s) chosen among the compounds belonging to the following groups : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

Growth hormone or its derivatives or any factor inducing growth hormone release and one or several compound(s) chosen among the compounds belonging to the following groups : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies can be administered simultaneously or at different times and/or at the same site or at different site(s) and/or in the same or in a different composition or medicament.

The cytokine group can comprise IL-1, IL-3, G-CSF, GM-CSF or SCF. The chemokine group can comprise MIP-1 α or thrombopoietin (TPO). The monoclonal antibody group can comprise anti-VLA-4 antibodies.

Preferably, Growth hormone or its derivatives or any factor inducing growth hormone is present in a composition which comprises granulocyte colony stimulating factor (G-CSF).

Preferably, Growth hormone or its derivatives or any factor inducing growth hormone is associated with G-CSF.

Growth hormone or its derivatives or any factor inducing growth hormone and G-CSF can be administered simultaneously or at different times and/or at the same site or at different site(s) and/or in the same or in a different composition or medicament.

Growth hormone or its derivatives or any factor inducing growth hormone release and G-CSF may advantageously be administered separately.

G-CSF may advantageously be administered in an amount comprised between 3 and 15 μg per kilogram of body weight, more particularly between 4 and 12 μg per kilogram of body weight.

A preferred dosage of G-CSF to be administered is around 5 μg or around 10 μg per kilogram of body weight.

In a preferred embodiment, Growth hormone or one of its derivatives is administered in an amount comprised between 20 and 50 μg per kilogram of body weight, more particularly between 30 and 40 μg per kilogram of body weight and G-CSF is administered in an amount comprised between 3 and 15 μg per kilogram of body weight, more particularly between 4 and 12 μg per kilogram of body weight.

In a preferred embodiment, Growth hormone or one of its derivatives is administered in an amount of 33 μg per kilogram of body weight and G-CSF is administered in an amount of around 5 μg or around 10 μg per kilogram of body weight.

According to the invention, the expression « administration in an amount sufficient to increase the number of circulating cells capable of regenerating hematopoiesis in vivo or to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo » can mean one or several administration(s), one or several times a day and during one or several days for a cumulated amount sufficient to increase the number of circulating cells capable of regenerating hematopoiesis in vivo or to reduce the volume of blood required to be processed in order to obtain the specified target number of

circulating cells capable of regenerating hematopoiesis in vivo.

The pharmaceutical compositions or compositions which are used in the methods and uses of the invention are in a pharmaceutical acceptable form optionally combined with an acceptable carrier.

These compositions can be administered by any means that achieve their intended purposes.

The compositions used in the methods or uses of the invention may be administered alone or in conjunction with other therapeutics directed to a disease or directed to other symptoms thereof.

The compositions used in the methods or uses of the invention may be administered by the intravenous or the subcutaneous route.

After intravenous administration, the elimination of hGH is described by first-order kinetics with a serum half-life of 12- 30 minutes in both animals and humans (Moore et al., 1988; Hendricks et al., 1985). Traditionally, intramuscular injection has been the method of choice as the preferred route of delivery. In humans, absorption of exogenous hGH appears to be more rapid from the intramuscular site, with a time to maximum concentration of two to three hours, compared to four to six hours after subcutaneous administration. The disappearance phase from serum has been reported to range from 12-20 hours for intramuscular administration, and 20-24 hours after subcutaneous administration (Albertsson-Wikland et al., 1986; Jorgensen et al., 1987).

The compositions used in the methods or uses of the invention may be administered by parenteral routes such as subcutaneous, intravenous, intramuscular, intraperitoneal, or transdermal route or by mucosal routes such as buccal or oral route.

If a method or use of the invention comprises the administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF, growth hormone or one of its derivatives or any factor inducing growth hormone release is preferably

administered three times a day and G-CSF is preferably administered daily.

The composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be a daily administration that can start up to 20 days pre-leukapheresis.

The composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered over a period of 5 days or over a period of 10 days, until leukapheresis or until the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo) is reached.

Preferably, the composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release is administered until leukapheresis and/or the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo) is reached.

Methods and uses of the invention are advantageously carried out after chemotherapy, radiotherapy, myelosuppressive therapy, transplantation or engraftment of cells capable of regenerating hematopoiesis in vivo or transplantation of bone-marrow.

Methods and uses of the invention are advantageously carried out around 7 days after the beginning of a chemotherapeutic treatment or around 2 days after the end of a chemotherapeutic treatment.

In a preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF are administered until leukapheresis, until mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, until increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, until reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, and/or until reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo. In this preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release is preferably administered three times a day and G-CSF is preferably administered once a day.

Methods and uses of the invention may be combined with a prior treatment called « chemopriming ». The « chemopriming » regimens which may be used are :

- high-dose cyclophosphamide (4 g/m^2) for patients with breast cancer or multiple myeloma,
- ifosfamide, etoposide for patients with non-Hodgkin's lymphoma or Hodgkin's disease,
- cyclophosphamide, etoposide, cisplatin (CVP) for patients with solid tumors (e.g., breast cancer).

To enhance the induction of cells capable of regenerating hematopoiesis in vivo rebound, methods and uses of the invention are started shortly after completion of chemopriming treatment and continued until completion of apheresis (5 to 12 $\mu\text{g/kg/d}$).

It is also noteworthy that in patients whose marrow stem cell pool is significantly diminished by prior chemotherapy, an additional chemopriming regimen might impair rather than induce cells capable of regenerating hematopoiesis in vivo peripheralization. Stem cells toxic chemotherapeutic agents such as busulfan, doxorubicin, melphalan, thiotepe and possibly fludarabine (and others) should not be part of a chemopriming regimen. On the other hand, cyclophosphamide is considered the ideal chemopriming drug with the least cells capable of regenerating hematopoiesis in vivo toxicity, although cardiotoxicity (dose > 4g/m²) and hemorrhagic cystitis are the well-known dose-limiting extramedullary side effects (Shepperd et al., 1990).

The population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo obtained from the peripheral blood by the methods and uses of the invention can be re-infused, grafted or transplanted into the same individual which is in this case the donor (autologous transplantation) or into different individuals (nonautologous transplantation).

The population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo obtained from the peripheral blood by the methods and uses of the invention are advantageously infused into an individual which has previously received one or several chemotherapy, radiotherapy, myelosuppressive, myeloablative or myelotoxic therapy.

Said operation of re-infusion, engraftment or transplantation belongs to the so-called Hematopoietic

Stem Cells Transplantation (HSCT) procedures. HSCT is a clinical procedure in which cells capable of regenerating hematopoiesis in vivo, obtained from bone marrow or peripheral blood, are transplanted to a patient.

An autologous transplantation is a transplantation in which donor and recipient are the same individual whereas a nonautologous transplantation is a transplantation in which donor and recipient are different individuals. The method of the invention encompasses both autologous and non-autologous transplantation.

In another part, the invention provides a method or a use of growth hormone or one of its derivatives or any factor inducing growth hormone release to enhance the mobilization or peripheralisation effect of G-CSF.

The invention provides a method or a use of growth hormone or one of its derivatives or any factor inducing growth hormone release to enhance mobilisation of circulating cells capable of regenerating hematopoiesis in vivo by G-CSF, to enhance increase the number of circulating cells capable of regenerating hematopoiesis in vivo by G-CSF, to enhance reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation by G-CSF, and/or to enhance reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo by G-CSF.

Thus, the administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF enhances or increases synergistically the mobilisation of circulating cells capable of regenerating hematopoiesis in vivo, enhances or increases synergistically the number of circulating cells capable of regenerating hematopoiesis in vivo, reduces the number

of leukapheresis required to collect sufficient amount of circulating cells for transplantation, and/or reduces the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo with respect to the effect(s) obtained by administering G-CSF alone or without growth hormone or one of its derivatives or any factor inducing growth hormone release.

The administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF allows to use lower dose(s) of G-CSF than if G-CSF is used alone or without growth hormone or one of its derivatives or any factor inducing growth hormone release.

The administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF can be carried out simultaneously or at different times and/or at the same site or at different site(s) and/or in the same or in a different composition or medicament.

In a second part, the invention provides new uses for enhancing hematopoietic reconstitution. The invention relates to an agent capable of promoting, enhancing or accelerating the hematopoietic regeneration, recovery or reconstitution. The invention provides new uses for enhancing hematopoietic reconstitution.

Thus, the invention relates to the use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament for enhancing hematopoietic reconstitution, in a human being.

Throughout the application, the term « enhancing » and all terms having the same root may be replaced by the term « promoting » or by the term « accelerating ».

Throughout the application, the term « reconstitution » and all terms having the same root may be replaced by the term « recovery » or by the term « regeneration ».

In another embodiment, the invention relates to the use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament for enhancing hematopoietic reconstitution following bone marrow transplantation in a human being.

In another embodiment, the invention relates to the use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament for enhancing engraftment of bone marrow or cells capable of regenerating hematopoiesis in vivo in a human being.

Another embodiment of the invention is the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for enhancing hematopoietic reconstitution after transplantation of cells capable of regenerating hematopoiesis in vivo.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for enhancing engraftment of cells capable of regenerating hematopoiesis in vivo.

Growth hormone may advantageously be human growth hormone.

Growth hormone and its derivatives may correspond to growth hormone and its derivatives which are recited above in this application in connection with the first part of the invention.

The hematopoietic reconstitution or the enhanced engraftment may be detected by an increase of the

Publ. No. 2136350

peripheral White Blood Cell (WBC) count and/or granulocytes count and/or lymphocytes count and/or platelet count and/or erythrocyte count.

An increase of the peripheral White Blood Cell (WBC) count and/or granulocytes count and/or lymphocytes count and/or platelet count and/or erythrocyte count may be detected by comparison with the rate of increase of said counts in an individual who has received the same transplantation regimen but who has not received any hematopoietic reconstitution treatment.

The hematopoietic reconstitution or the enhanced engraftment may be detected by reduction of the period of time necessary to recover a normal or standard peripheral White Blood Cell (WBC) count and/or granulocytes count and/or neutrophil count and/or lymphocytes count and/or platelet count and/or erythrocyte count.

A normal or standard peripheral White Blood Cell (WBC) count and/or granulocytes count and/or neutrophil count and/or lymphocytes count and/or platelet count and/or erythrocyte count is the one which is measured in a healthy individual or in an individual which has not received any myeloablative, myelotoxic or myelosuppressive therapy, any chemotherapy, any radiotherapy or any transplantation.

A normal neutrophil count may be at least 0.5×10^9 neutrophil cells per liter of peripheral blood.

A normal platelet count may be at least 20×10^9 per liter of peripheral blood.

The hematopoietic reconstitution or enhancing engraftment may be detected by a reduction of the extent and/or duration of neutropenia and/or thrombocytopenia and/or anemia and/or hemorrhages and/or duration of prophylaxis.

The hematopoietic reconstitution or the enhancing engraftment may be detected by a reduction of the duration and/or severity of fever and/or infections.

A reduction of the extent and/or duration of neutropenia and/or thrombocytopenia and/or anemia and/or hemorrhages and/or duration of prophylaxis or a reduction of the duration and/or severity of fever and/or infections may be compared with said extent and/or duration and/or severity measured in an individual who has received the same transplantation regimen, the same chemotherapy, the same radiotherapy or the same myelosuppressive, myeloablative or myelotoxic therapy but who has not received any hematopoietic reconstitution treatment.

The hematopoietic reconstitution or the enhancing engraftment may be detected by a recovery of granulocytes which is at least 1000 per microliter of peripheral blood.

The hematopoietic reconstitution or the enhancing engraftment may be detected by a recovery of platelet count which is at least 50,000 per microliter of peripheral blood.

In another embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for treating a neoplastic disease, an hematological disorder, malignancies, Severe Combined Immune Deficiencies (SCIDs), congenitally or genetically determined hematopoietic abnormalities, anemia, aplastic anemia, leukemia and/or osteopetrosis.

A neoplastic disease may be breast cancer.

In another embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for reducing the bone marrow aplasia period

which follows transplantation, chemotherapy, radiotherapy or myeloablative, myelosuppressive or myelotoxic therapy, for preventing or treating opportunistic infections after transplantation, chemotherapy, radiotherapy or myeloablative, myelosuppressive or myelotoxic therapy or for limiting the risk of tumor recurrence after transplantation, chemotherapy, radiotherapy or myeloablative, myelosuppressive or myelotoxic therapy.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating secondary effects of myeloablative, myelotoxic or myelosuppressive therapy and/or radiotherapy and/or chemotherapy and/or transplantation.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating neutropenia and/or thrombocytopenia.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating anemia following radiotherapy and/or chemotherapy and /or hematopoietic stem cells transplantation and/or transplantation of cells capable of regenerating hematopoiesis and/or Bone marrow transplantation and/or myelosuppressive or myelotoxic therapy.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating neutropenia, following radiotherapy and/or chemotherapy and /or hematopoietic stem cells transplantation and/or

transplantation of cells capable of regenerating hematopoiesis and/or Bone marrow transplantation and/or myelosuppressive or myelotoxic therapy.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating thrombocytopenia, following radiotherapy and/or chemotherapy and /or hematopoietic stem cells transplantation and/or transplantation of cells capable of regenerating hematopoiesis and/or Bone marrow transplantation and/or myelosuppressive or myelotoxic therapy.

The cells capable of regenerating hematopoiesis in vivo may belong to one or several of the following groups of cells : CD34⁺ cells, CD34⁺CD33⁺ cells, CD34⁺CD38⁻ cells, CD34⁺Thy-I cells, CD34⁺Thy-ICD38⁻ cells, CD33⁺ cells, stem cells, progenitor cells, long-term culture initiating cells (LTC-IC), cells that fulfill self renewal potential, cells that fulfill pluripotential characteristics, cells that initiate long term bone marrow culture.

Determination of amounts of growth hormone, of one of its derivatives or of any factor inducing growth hormone release to be administered in a method or use of the invention described above is within the skill of the art.

Typical dosage of growth hormone, of one of its derivatives or of any factor inducing growth hormone release will start at about 1 microgram per kilogram of the patient weight per day and dose will be escalated until the desired effect (hematopoietic recovery or engraftment) is reached.

The dosage of growth hormone, of one of its derivatives or of any factor inducing growth hormone release administered depends upon the age, sex, health

and weight of the donor, type of previous or concurrent treatment, if any, frequency of the treatment and the nature of the effect desired.

Growth hormone or its derivatives or any factor inducing growth hormone release may be administered alone or in conjunction or association with other factors.

Growth hormone or its derivatives or any factor inducing growth hormone release may advantageously be present in a composition or a medicament which comprises further one or several compound(s) chosen among the compounds belonging to the following groups : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

Growth hormone or its derivatives or any factor inducing growth hormone release and one or several compound(s) chosen among the compounds belonging to the following groups : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies can be administered simultaneously or at different times and/or at the same site or at different site(s) and/or in the same or in a different composition or medicament.

The cytokine group can comprise IL-1, IL-3, G-CSF, GM-CSF or SCF. The chemokine group can comprise MIP-1 α or thrombopoietin (TPO). The monoclonal antibody group can comprise anti-VLA-4 antibodies.

Preferably, Growth hormone or its derivatives or any factor inducing growth hormone is present in a composition or a medicament which comprises granulocyte colony stimulating factor (G-CSF).

Preferably, Growth hormone or its derivatives or any factor inducing growth hormone is associated with G-CSF.

Growth hormone or its derivatives or any factor inducing growth hormone and G-CSF can be administered simultaneously or at different times and/or at the same

site or at different site(s) and/or in the same or in a different composition or medicament.

Growth hormone or its derivatives or any factor inducing growth hormone release and G-CSF may advantageously be administered separately.

Growth hormone or its derivatives or any factor inducing growth hormone release and/or G-CSF are administered in an amount sufficient to enhance hematopoietic reconstitution or engraftment.

Administration in an amount sufficient to enhance hematopoietic reconstitution or engraftment can mean one or several administration(s), one or several times a day and during one or several days for a cumulated amount sufficient to enhance hematopoietic reconstitution or engraftment.

The pharmaceutical compositions or medicaments or compositions which are used in the methods and uses of the invention are in a pharmaceutical acceptable form optionally combined with an acceptable carrier.

These compositions or medicaments can be administered by any means that achieve their intended purposes.

The compositions or medicaments used in the methods or uses of the invention may be administered alone or in conjunction with other therapeutics directed to a disease or directed to other symptoms thereof.

The compositions or medicaments used in the methods or uses of the invention may be administered by the intravenous or the subcutaneous route.

The compositions or medicaments used in the methods or uses of the invention may be administered by parenteral routes such as subcutaneous, intravenous, intramuscular, intraperitoneal, or transdermal route or by mucosal routes such as buccal or oral route.

The composition or medicament comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered by parenteral routes such as subcutaneous, intravenous, intramuscular, intraperitoneal, or transdermal route or by mucosal routes such as buccal or oral route.

Preferably, growth hormone or one of its derivatives or any factor inducing growth hormone release is administered subcutaneously.

The total dose or amount required for each treatment, method or use of the invention may be administered in multiple or single dose.

The composition or medicament comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered daily or three times a day.

Preferably, the composition or medicament comprising growth hormone or one of its derivatives or any factor inducing growth hormone release is administered three times a day.

Preferably, growth hormone or one of its derivatives or any factor inducing growth hormone release is administered daily or three times a day.

In a preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release is administered three times a day.

If a method or use of the invention comprises the administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF, G-CSF is preferably administered once a day and/or subcutaneously.

If a method or use of the invention comprises the administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF, growth hormone or one of its derivatives or

any factor inducing growth hormone release is preferably administered three times a day and G-CSF is preferably administered daily.

The administration of the medicament may be made over a period of 3 days along, until leukapheresis or until full recovery.

The administration of the medicament may be made from day 1 to day 3 after transplantation.

The term « transplantation » encompasses bone marrow transplantation or hematopoietic stem cells transplantation.

The composition or medicament comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be a daily administration that can start up to 20 days pre-leukapheresis.

The composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered over a period of 5 days or over a period of 10 days until the desired effect (hematopoietic recovery or engraftment) is reached.

Preferably, the composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release is administered until the desired effect (hematopoietic recovery or engraftment) is reached.

Methods and uses of the invention are advantageously carried out after chemotherapy, radiotherapy, myelosuppressive therapy, transplantation or engraftment of cells capable of regenerating hematopoiesis in vivo or transplantation of bone-marrow.

Methods and uses of the invention are advantageously carried out around 7 days after the beginning of a chemotherapeutic treatment or around 2 days after the end of a chemotherapeutic treatment.

In a preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF are administered until hematopoietic reconstitution or engraftment. In this preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release is preferably administered three times a day and G-CSF is preferably administered once a day.

Growth hormone used in the medicament may advantageously be recombinant growth hormone.

Growth hormone used in the medicament may advantageously be human growth hormone.

In a third part, the invention provides a combination of the methods and uses of the first part of the invention (mobilisation) with the methods and uses of the second part of the invention (recovery).

Said combination methods and uses are mobilisation and recovery methods which may be applied in cases autologous transplantation of hematopoietic stem cells wherein the donor and the recipient is the same person or individual. Thus, growth hormone or one of its derivatives or any factor inducing growth hormone release can be used as a mobilising agent in a first mobilisation step which is a pre-treatment in view of blood cells extraction and as an hematopoietic recovery agent in a second step following transplantation.

Said combination methods and uses are very useful. In fact, transplantation of cells mobilized by growth hormone or one of its derivatives or any factor inducing growth hormone release to a patient results in faster haematological recovery than transplantation without a prior mobilization treatment of said patient.

Methods and uses of the invention can be applied in many clinically important fields, namely autologous bone marrow transplantation, allogeneic bone marrow

Methods and uses of the invention can be applied to treat a patient who has received radiotherapy or chemotherapy, who has been transplanted with bone-marrow or cells capable of regenerating hematopoiesis in vivo, or who has received myelotoxic or myeloablative therapy.

Methods and uses of the invention can be applied to treat a patient who has received radiotherapy or chemotherapy, who has been transplanted with bone-marrow or cells capable of regenerating hematopoiesis in vivo, or who has received myelotoxic or myeloablative therapy.

LEGEND OF THE FIGURES :

Figure 1 :Abbreviations :

- GH : Growth Hormone
- G-CSF : Granulocyte-Cell Stimulating Factor
- ND : Not Detectable

Figure 2 :

This graph depicts the number of CD34⁺ cells/ μ l of blood obtained in a patient during 3 cycles of chemotherapy after a mobilization treatment with G-CSF alone (cycle 1), GH + G-CSF (cycle 2) and G-CSF alone (cycle 3).

ExamplesAbbreviations and notes :

- BFU-E : burst forming unit, erythroid
- CFU-C : colony forming unit, culture
- CFU-GM : colony forming unit, granulocyte and macrophage
- CFU-Meg : colony forming unit, megakaryocyte
- G-CSF : granulocyte colony stimulating factor
- IGF-I : insulin growth factor I
- LTC-IC : long term culture initiating cell
- HGH : human growth hormone

50

- RhG-CSF : recombinant human granulocyte colony stimulating factor
- RhGH : recombinant human growth hormone

Example 1 : Mobilization activity of hGH studied in a murine preclinical model

BALB/c mice are given 10 μ L intraperitoneal injections of rhGH every day for total of 10 days. The total CFU-C or BFU-E activity circulating in the peripheral blood on day 5 and day 10, respectively, is determined according to standard in vitro culture techniques, and compared with :

- (i) steady-state pretreatment levels,
- (ii) absolute CFU-C and BFU-E counts on day 3 and day 5, respectively following treatment with rhG-CSF given intraperitoneally at 10 μ L every day for 5 consecutive days.

Example 2 : Selection criteria for the mobilization and recovery clinical studies

A) Inclusion criteria :

- Written informed consent
- Age 18 years and 60 years
- Histologically confirmed high-risk cancer (lymphome cancer) undergoing high-dose chemotherapy according to current INT guidelines.

B) Exclusion criteria :

- Patients heavily pretreated with chemotherapy courses of chemotherapy) and/or radiotherapy.

51

- Renal (creatinine > 1,5 N), or hepatic insufficiency and/or SGPT > 2,5 N ; bilirubin > 1,5 N), or severe CNS or psychiatric disease.
- Clinically significant cardiac disease or myocardial. Left ventricular ejection fraction < 50% at rest by echocardiography assessment or < 55% by isotopic measurement.
- Hepatitis B or C, or HIV test positive.

Example 3 : Baseline study procedure for the mobilization and recovery clinical studies

Several parameters are studied during the mobilisation and recovery clinical studies :

- Complete medical history, physical examination, cardiac examination, left ventricular ejection fraction (LVEF) by multigated scintigraphic scan or echography, chest X-ray
- Pregnancy test (if applicable)
- HBV, HCV and HIV test
- Complete blood count with differential
- Absolute counts of circulating CD34⁺ cells and CFU
- Blood chemistry (transaminases, serum phosphatase, gammaGT, LDH, total bilirubin, BUN, creatinine, glycemia, Na, K, Ca, P, uric acid, total protein, albumin, cholesterol, triglycerides
- Bilateral bone marrow biopsy
- Informed consent

Example 4 : Main parameters of toxicity for the mobilization and recovery clinical studies

- Tumor growth (mobilization study only)
- Clinical and instrumental symptoms
- Laboratory tests for cardiac, liver and renal function

Example 5 : Mobilization clinical study

A) Objectives of the mobilization clinical study

- To assess the activity of rhGH in :
 - (i) increasing circulating CD34⁺ cells, and
 - (ii) expanding the bone marrow hematopoietic compartment, so to allow an enhanced mobilization by subsequent rhG-CSF administration
- To assess the safety and tolerability of rhGH, given with rhG-CSF to cancer patients following chemotherapy (hematologic recovery study).

B) Treatment plan

Mobilization study with rhGH :

- rhGH is administered from day 1 to 10 by the intravenous route. Dosage of rhGH is started at about 1 micrograms per kilogram of the patient weight per day and dose will be escalated until the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase in said donor of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating

53

cells capable of regenerating hematopoiesis in vivo) is reached.xx $\mu\text{g/kg}$ QD, iv)

Mobilization study with rhGH and rhG-CSF :

- rhGH administration : rhGH is administered from day 1 to 5 by the intravenous route. Dosage of rhGH is started at about 1 micrograms per kilogram of the patient weight per day and dose will be escalated until the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase in said donor of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo) is reached.
- rhG-CSF administration ($10 \mu\text{g/kg}$ QD, iv) from completion of CD34⁺ cell harvest (target cell dose is 8×10^6 CD34⁺ cells/kg body weight).

C) Main parameters of activity

Starting from day +6, the following parameters are assessed :

- Absolute CD34⁺ cell counts/ μL (daily in the periphery and once in the leukapheresed cells)
- Absolute CFU-GM counts/ μL (daily in the periphery and once in the leukapheresed cells)

D) Study procedure

- Daily assessment of CD34⁺ cells/ μL and CFU-GM peripheral blood, from day +5 until leukapheresis.
- Total yield of CD34⁺ cells, CFU-GM, BFU-E, CFU IC in leukapheresed cells.

- Toxicity assess through clinical and in examinations (EKG, chest X-ray, and other examinations as required).
- Measurement and evaluation of all tumor parameters ending the mobilization study

Example 6 : Recovery clinical study

A) Objectives of the recovery clinical study

- To assess the ability of rhGH, given alone or in combination to hasten the recovery of WBC, RBC and platelets in the peripheral blood of cancer patients treated with high-dose chemotherapy and peripheral blood stem-cell autografting.
- To assess the safety and tolerability of rhGH, given with rhG-CSF to cancer patients following chemotherapy.

B) Treatment plan

- Administration of high-dose chemotherapy, followed by infusion on day 0 of an optimal amount (i.e. 8×10^6 CD34⁺ cells/kg) of crypreserved cells harvested in the mobilization study.
- Co-administration (iv) of rhGH and rhG-CSF $\mu\text{g/kg}$ QD, iv) from day 1 until stable (i.e. for three consecutive days) recovery of granulocytes above $1000 \mu\text{L}$, and platelet counts above $50,000/\mu\text{L}$.

C) Main parameters of activity

Starting from day +0, and until full and stable recovery, the following parameters will be assessed :

- Absolute granulocyte counts/ μL (daily)
- Absolute platelet counts/ μL (daily)
- Absolute erythrocyte counts/ μL (daily)
- Granulocyte nadir

- Platelet nadir
- Extent and duration of neutropenia
- Extent and duration of thrombocytopenia
- Extent and duration of hematopoietic support transfusions, RBC transfusions)
- Duration of infectious prophylaxis, and infections
- Hemorrhages

D) Study procedure

- Daily assessment of WBC, RBC and platelet count
- Number of platelet transfusions
- Number of RBC transfusions
- Type and severity of fever and documented infection
- Clinical and instrumental assessment of toxicities hematological

Example 7 : Results of the mobilization clinical studies

I - Studies of mobilization within 2 cycles of chemotherapy

A) Treatment plan

Three patients with relapsed Hodgkin's disease have received the following two cycles of treatment :

- cycle 1(control cycle) :
 - Ifosfamide (agent for chemotherapy) : 3 g/m² iv (intravenous) (once a day), day 1-4 ;
 - Vinorelbine (agent for chemotherapy) : 25 mg/m² iv (once a day), day 1 and 5 ;
 - G-CSF : 5 µg/kg sc (subcutaneous) (once a day), from day 7 until leukapheresis or until recovery of a sufficient number of CD34+ cells(around 3 to 8-10⁶ cells/kg).

1. The first part of the document is a list of references. The references are listed in a standard format, with the author's name, the title of the work, and the publisher. The references are as follows:

The results are depicted in the table of figure 1.

As compared with the control (cycle 1), the addition of rhGH in cycle 2 resulted in :

- The increase of the number of CD34⁺ leukapheresed cells induced by GH allows the harvest from all three patients of an amount of CD34⁺ cells adequate for autologous transplantation (around 3 to 8-10⁶ cells/kg).

II - Studies of mobilization within 3 cycles of chemotherapy

A) Treatment plan

One patient with relapsed Hodgkin's disease has received the following 3 cycles of treatment :

- cycle 1 :

- Ifosfamide (agent for chemotherapy) : 3 g/m² iv (intravenous) (once a day), day 1-4 ;
- Vinorelbine (agent for chemotherapy) : 25 mg/m² iv (once a day), day 1 and 5 ;
- G-CSF : 5 µg/kg sc (subcutaneous) (once a day), from day 7 until leukapheresis or recovery of a sufficient amount of CD34+ cells (around 3 to 8-10⁶ cells/kg).

- cycle 2 :

- Ifosfamide : 3 g/m² iv (once a day), day 1-4 ;
- Vinorelbine : 25 mg/m² iv (once a day), day 1 and 5 ;
- G-CSF : 5 µg/kg sc (once a day) from day 7 until recovery of a sufficient amount of CD34+ cells (around 3 to 8-10⁶ cells/kg) or until leukapheresis ;
- rhGH (recombinant human Growth Hormone) : 33 µg/kg sc (three times a day), from day 7 until recovery of a sufficient amount of CD34+ cells (around 3 to 8-10⁶ cells/kg) or until leukapheresis.

- cycle 3 :

- Ifosfamide (agent for chemotherapy) : 3 g/m² iv (intravenous) (once a day), day 1-4 ;
- Vinorelbine (agent for chemotherapy) : 25 mg/m² iv (once a day), day 1 and 5 ;
- G-CSF : 5 µg/kg sc (subcutaneous) (once a day), from day 7 until leukapheresis or until recovery of a

58

The results of the clinical treatment recited in section A above are depicted in the graph of Figure 2.

Graph of figure 2 shows the time course of CD34⁺ cell mobilization, following three consecutive chemotherapy cycles beginning every 21 days.

Each point depicted in the graph of figure 2 corresponds to the measure of the number of CD34⁺ cells/ μ l of blood found in a sample of blood of 1 milliliter.

The results show that cycle 2 (addition of rhGH) is clearly superior to cycles 1 and 3. Thus, the mobilization of CD34⁺ in the blood is enhanced by the addition of rhGH.

Enhancement of mobilization of CD34⁺ cells in the blood by GH is high, especially since the patient studied receives several courses of myelotoxic chemotherapy, and since each subsequent course hampers the extent of mobilization. The declining numbers of circulating CD34⁺ cells after consecutive myelotoxic chemotherapy and mobilization cycles can be observed by comparing cycle 1 and cycle 3.

The blood of the patient is leukapheresed when the number of CD34⁺ cells/ μ l of blood measured is maximal (day 13 of cycle 1 ; day 20 of cycle 2).

[illegible]

The leukapheresed cells are cryopreserved and will be reinfused in the patient after a myeloablative therapy.

WO 00/40260

- Adelman, J.P., Hayflick, J.S., Vasser, M., and Seeburg, P.H. (1983). In vitro deletional mutagenesis for bacterial production of the 20,000- dalton form of human pituitary growth hormone. DNA 2, 183-193.
- Albertsson-Wikland, K., Westphal, O., and Westgren, U. (1986). Daily subcutaneous administration of human growth hormone in growth hormone deficient children. Acta Paediatr.Scand. 75, 89-97.
- Alexander, W.S. (1998). Cytokines in hematopoiesis. Int.Rev.Immunol. 16, 651-682.
- Anderlini, P. and Korbling, M. (1997). The use of mobilized peripheral blood stem cells from normal donors for allografting. Stem.Cells 15, 9-17.
- Becker, G.W., Bowsher, R.R., Mackellar, W.C., Poor, M.L., Tackitt, P.M., and Riggan, R.M. (1987). Chemical, physical, and biological characterization of a dimeric form of biosynthetic human growth hormone. Biotechnol.Appl.Biochem. 9, 478-487.
- Becker, G.W., Tackitt, P.M., Bromer, W.W., Lefeber, D.S., and Riggan, R.M. (1988). Isolation and characterization of a sulfoxide and a desamido derivative of biosynthetic human growth hormone. Biotechnol.Appl.Biochem. 10, 326-337.
- Bewley, T.A. and Li, C.H. (1975).
The chemistry of human pituitary growth hormone. Adv.Enzymol.Relat.Areas.Mol.Biol. 42:73-166, 73-166.

Cunningham, B.C., Mulkerrin, M.G., and Wells, J.A. (1991). Dimerization of human growth hormone by zinc. Science 253, 545-548.

DeNoto, F.M., Moore, D.D., and Goodman, H.M. (1981). Human growth hormone DNA sequence and mRNA structure : possible alternative splicing. Nucleic.Acids.Res. 9, 3719-3730.

Derfalvi, B., Sallai, P., Nemet, K., Szalai, C., Kenesei, E., Tulassay, T., and Falus, A. (1998). [The in vitro effect of recombinant human growth hormone on lymphocyte and granulocyte function in healthy and uremic children]. Orv.Hetil. 139, 1847-1850.

Fischer, A., Haddad, E., Jabado, N., Casanova, J.L., Blanche, S., Le Deist, F., and Cavazzana-Calvo, M. (1998). Stem cell transplantation for immunodeficiency. Springer Semin.Immunopathol. 19, 479-492.

Geffner, M. (1997). Effects of growth hormone and insulin-like growth factor I on T- and B- lymphocytes and immune function. Acta Paediatr.Suppl. 423 :76-9, 76-79.

Gertler, A., Shamay, A., Cohen, N., Ashkenazi, A., Friesen, H.G., Levanon, A., Gorecki, M., Aviv, H., Hadary, D., and Vogel, T. (1986). Inhibition of lactogenic activities of ovine prolactin and human growth hormone (hGH) by a novel form of a modified recombinant hGH. Endocrinology 118, 720-726.

Gianni, A.M., Bregni, M., Siena, S., Villa, S., Sciorelli, G.A., Ravagnani, F., Pellegris, G., and Bonadonna, G. (1989). Rapid and complete hemopoietic reconstitution following combined transplantation of

Q. 10. 11. 12. 13. 14. 15. 16. 17. 18. 19. 20. 21. 22. 23. 24. 25. 26. 27. 28. 29. 30. 31. 32. 33. 34. 35. 36. 37. 38. 39. 40. 41. 42. 43. 44. 45. 46. 47. 48. 49. 50. 51. 52. 53. 54. 55. 56. 57. 58. 59. 60. 61. 62. 63. 64. 65. 66. 67. 68. 69. 70. 71. 72. 73. 74. 75. 76. 77. 78. 79. 80. 81. 82. 83. 84. 85. 86. 87. 88. 89. 90. 91. 92. 93. 94. 95. 96. 97. 98. 99. 100. 101. 102. 103. 104. 105. 106. 107. 108. 109. 110. 111. 112. 113. 114. 115. 116. 117. 118. 119. 120. 121. 122. 123. 124. 125. 126. 127. 128. 129. 130. 131. 132. 133. 134. 135. 136. 137. 138. 139. 140. 141. 142. 143. 144. 145. 146. 147. 148. 149. 150. 151. 152. 153. 154. 155. 156. 157. 158. 159. 160. 161. 162. 163. 164. 165. 166. 167. 168. 169. 170. 171. 172. 173. 174. 175. 176. 177. 178. 179. 180. 181. 182. 183. 184. 185. 186. 187. 188. 189. 190. 191. 192. 193. 194. 195. 196. 197. 198. 199. 200. 201. 202. 203. 204. 205. 206. 207. 208. 209. 210. 211. 212. 213. 214. 215. 216. 217. 218. 219. 220. 221. 222. 223. 224. 225. 226. 227. 228. 229. 230. 231. 232. 233. 234. 235. 236. 237. 238. 239. 240. 241. 242. 243. 244. 245. 246. 247. 248. 249. 250. 251. 252. 253. 254. 255. 256. 257. 258. 259. 260. 261. 262. 263. 264. 265. 266. 267. 268. 269. 270. 271. 272. 273. 274. 275. 276. 277. 278. 279. 280. 281. 282. 283. 284. 285. 286. 287. 288. 289. 290. 291. 292. 293. 294. 295. 296. 297. 298. 299. 300. 301. 302. 303. 304. 305. 306. 307. 308. 309. 310. 311. 312. 313. 314. 315. 316. 317. 318. 319. 320. 321. 322. 323. 324. 325. 326. 327. 328. 329. 330. 331. 332. 333. 334. 335. 336. 337. 338. 339. 340. 341. 342. 343. 344. 345. 346. 347. 348. 349. 350. 351. 352. 353. 354. 355. 356. 357. 358. 359. 360. 361. 362. 363. 364. 365. 366. 367. 368. 369. 370. 371. 372. 373. 374. 375. 376. 377. 378. 379. 380. 381. 382. 383. 384. 385. 386. 387. 388. 389. 390. 391. 392. 393. 394. 395. 396. 397. 398. 399. 400. 401. 402. 403. 404. 405. 406. 407. 408. 409. 410. 411. 412. 413. 414. 415. 416. 417. 418. 419. 420. 421. 422. 423. 424. 425. 426. 427. 428. 429. 430. 431. 432. 433. 434. 435. 436. 437. 438. 439. 440. 441. 442. 443. 444. 445. 446. 447. 448. 449. 450. 451. 452. 453. 454. 455. 456. 457. 458. 459. 460. 461. 462. 463. 464. 465. 466. 467. 468. 469. 470. 471. 472. 473. 474. 475. 476. 477. 478. 479. 480. 481. 482. 483. 484. 485. 486. 487. 488. 489. 490. 491. 492. 493. 494. 495. 496. 497. 498. 499. 500. 501. 502. 503. 504. 505. 506. 507. 508. 509. 510. 511. 512. 513. 514. 515. 516. 517. 518. 519. 520. 521. 522. 523. 524. 525. 526. 527. 528. 529. 530. 531. 532. 533. 534. 535. 536. 537. 538. 539. 540. 541. 542. 543. 544. 545. 546. 547. 548. 549. 550. 551. 552. 553. 554. 555. 556. 557. 558. 559. 560. 561. 562. 563. 564. 565. 566. 567. 568. 569. 570. 571. 572. 573. 574. 575. 576. 577. 578. 579. 580. 581. 582. 583. 584. 585. 586. 587. 588. 589. 590. 591. 592. 593. 594. 595. 596. 597. 598. 599. 600. 601. 602. 603. 604. 605. 606. 607. 608. 609. 610. 611. 612. 613. 614. 615. 616. 617. 618. 619. 620. 621. 622. 623. 624. 625. 626. 627. 628. 629. 630. 631. 632. 633. 634. 635. 636. 637. 638. 639. 640. 641. 642. 643. 644. 645. 646. 647. 648. 649. 650. 651. 652. 653. 654. 655. 656. 657. 658. 659. 660. 661. 662. 663. 664. 665. 666. 667. 668. 669. 670. 671. 672. 673. 674. 675. 676. 677. 678. 679. 680. 681. 682. 683. 684. 685. 686. 687. 688. 689. 690. 691. 692. 693. 694. 695. 696. 697. 698. 699. 700. 701. 702. 703. 704. 705. 706. 707. 708. 709. 710. 711. 712. 713. 714. 715. 716. 717. 718. 719. 720. 721. 722. 723. 724. 725. 726. 727. 728. 729. 730. 731. 732. 733. 734. 735. 736. 737. 738. 739. 740. 741. 742. 743. 744. 745. 746. 747. 748. 749. 750. 751. 752. 753. 754. 755. 756. 757. 758. 759. 760. 761. 762. 763. 764. 765. 766. 767. 768. 769. 770. 771. 772. 773. 774. 775. 776. 777. 778. 779. 780. 781. 782. 783. 784. 785. 786. 787. 788. 789. 790. 791. 792. 793. 794. 795. 796. 797. 798. 799. 800. 801. 802. 803. 804. 805. 806. 807. 808. 809. 810. 811. 812. 813. 814. 815. 816. 817. 818. 819. 820. 821. 822. 823. 824. 825. 826. 827. 828. 829. 830. 831. 832. 833. 834. 835. 836. 837. 838. 839. 840. 841. 842. 843. 844. 845. 8

Heather J. et al. Blood 74 ; 1563-1570 (1989).

Henry, D. (1997). Haematological toxicities associated with dose-intensive chemotherapy, the role for and use of recombinant growth factors. Ann.Oncol. 8 Suppl 3:S7-10, S7-10.

Jorgensen, K.D., Monrad, J.D., Brondum, L., and Dinesen, B. (1988). Pharmacokinetics of biosynthetic and pituitary human growth hormones in rats. Pharmacol.Toxicol. 63, 129-134.

Lewis, U.J., Peterson, S.M., Bonewald, L.F., Seavey, B.K., and VanderLaan, W.P. (1977). An interchain disulfide dimer of human growth hormone. J.Biol.Chem. 252, 3697-3702.

Lewis, U.J., Singh, R.N., Bonewald, L.F., and Seavey, B.K. (1981). Altered proteolytic cleavage of human growth hormone as a result of deamidation. J.Biol.Chem. 256, 11645-11650.

Merchav, S., Tatarsky, I., and Hochberg, Z. (1988). Enhancement of erythropoiesis in vitro by human growth hormone is mediated by insulin-like growth factor I. Br.J.Haematol. 70, 267-271.

Moore, J.A., Rudman, C.G., MacLachlan, N.J., Fuller, G.B., Burnett, B., and Frane, J.W. (1988). Equivalent potency and pharmacokinetics of recombinant human growth hormones with or without an N-terminal methionine. *Endocrinology* 122, 2920-2926.

Siena et al. Blood 74 ; 1905-1914 (1989)

Siena et al. Blood 77 ; 400-409 (1991)

Singh, R.N., Seavey, B.K., Rice, V.P., Lindsey, T.T., and Lewis, U.J. (1974). Modified forms of human growth hormone with increased biological activities. *Endocrinology* 94, 883-891.

Stolar, M.W., Amburn, K., and Baumann, G. (1984). Plasma "big" and "big-big" growth hormone (GH) in man : an oligomeric series composed of structurally diverse GH monomers. J.Clin.Endocrinol.Metab. 59, 212-218.

Stolar, M.W. and Baumann, G. (1986). Big growth hormone forms in human plasma : immunochemical evidence for their pituitary origin. *Metabolism* 35, 75-77.

Thorlacius-Ussing, O. (1987). Zinc in the anterior pituitary of rat : a histochemical and analytical work. *Neuroendocrinology*. 45, 233-242.

Tian, Z.G., Woody, M.A., Sun, R., Welniak, L.A., Raziuddin, A., Funakoshi, S., Tsarfaty, G., Longo, D.L., and Murphy, W.J. (1998). Recombinant human growth hormone promotes hematopoietic reconstitution after syngeneic bone marrow transplantation in mice. *Stem.Cells* 16, 193-199.

Valerio, G., Di Maio, S., Salerno, M., Argenziano, A., Badolato, R., and Tenore, A. (1997). Assessment of red blood cell indices in growth-hormone-treated children. *Horm.Res.* 47, 62-66.

Van Hoef, M.E. (1998). Haematological recovery after high-dose consolidation chemotherapy with peripheral blood progenitor cell rescue: the effects of the mobilization regimen and post-transplant growth factors. *Neth.J.Med.* 52, 30-39.

Vihervuori, E., Virtanen, M., Koistinen, H., Koistinen, R., Seppala, M., and Siimes, M.A. (1996). Hemoglobin level is linked to growth hormone-dependent proteins in short children. *Blood* 87, 2075-2081.

Waters, T.M., Bennett, C.L., Pajean, T.S., Sobocinski, K.A., Klein, J.P., Rowlings, P.A., and Horowitz, M.M. (1998). Economic analyses of bone marrow and blood stem cell transplantation for leukemias and lymphoma : what do we know ? Bone Marrow Transplant. 21, 641-650.

Weaver, A. and Testa, N.G. (1998). Stem cell factor leads to reduced blood processing during apheresis or the use of whole blood aliquots to support dose-intensive chemotherapy. Bone Marrow Transplant. 22, 33-38.

CLAIMS

1. Method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising:

- a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo.
- b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.
- c) re-infusing, grafting or transplanting said isolated population into the same individual or into different individuals.

2. Method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising:

- a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release, simultaneously or separately with a composition comprising G-CSF, in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo.
- b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

3. Method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising:

a) administering to a donor, over a period of up to 20 days, a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo.

b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

4. Method according to any one of claims 1 to 3 wherein the circulating cells capable of regenerating hematopoiesis in vivo are CD34⁺ cells.

5. Method according to claim 4 wherein the increased number of CD34⁺ cells is more than 10, 25, 34 or 80 CD34⁺ cells per microliter of donor peripheral blood.

6. Method according to claim 4 wherein the increased number of CD34⁺ cells is at least 2×10^6 , 4×10^6 , 5×10^6 , 6×10^6 , 8×10^6 , 15×10^6 CD34⁺ cells per kilogram of donor body weight.

7. Method according to any one of claims 1 to 3 wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to around or more than 500 CFU-GM per milliliter of donor peripheral blood.

8. Method according to any one of claims 1 to 3 wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to an increased level of CFU-C, CFU-Meg or BFU-E in donor peripheral blood.

9. Method according to any one of claims 1 to 3 wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo substantially corresponds to a white blood cell count of around or more than 1000 cells per microliter of donor peripheral blood.

10. Method according to any one of claims 1 to 3 wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to around or more than 1×10^5 GM-CFC per kilogram of donor or recipient body weight.

11. Method according to any one of claims 1 to 3 wherein the circulating cells capable of regenerating hematopoiesis in vivo are CD34⁺/CD33⁻ cells and/or CD34⁺/CD38⁻ cells and/or CD34⁺/Thy-I cells and/or CD34⁺/Thy-I/CD38⁻ cells and/or CD33⁺ cells and/or bone-marrow stem cells and/or progenitor cells and/or long-term culture initiating cells (LTC-IC) and/or cells that fulfill self renewal potential and/or cells that fulfill pluripotential characteristics and/or cells that initiate long term bone marrow culture and/or cells that can generate multiple cell lineages.

12. Method according to any one of claims 1 to 3 wherein the target number of circulating cells capable of regenerating hematopoiesis in vivo is at least 2×10^4 LTC-IC per kg of donor or recipient body, around or more than 2×10^6 CD34⁺ cells per kilogram of donor or recipient body weight, around or more than 4×10^6 CD34⁺ cells per kilogram of donor or recipient body weight or around or more than 8×10^6 CD34⁺ cells per kilogram of donor or recipient body weight.

13. Method according to any one of claims 1 to 3 wherein the volume of blood processed in step (b) is comprised in a range of about 30 to about 900 milliliters.

14. Method according to any one of claims 1 to 3 wherein the composition comprises one or several further compounds chosen among the following groups of compounds: hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

15. Method according to claim 1 or 3 wherein the composition is administered simultaneously or separately with one or more further compositions comprising one or several compounds chosen among the following groups of compounds: hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

16. Method according to claim 14 or 15 wherein the cytokine group comprises IL-1, IL-3, G-CSF, GM-CSF or SCF; the chemokine group comprises MIP-1 α or thrombopoietin (TPO); the monoclonal antibody group comprises anti-VLA-4 antibodies.

17. Method according to claim 14 or 15 wherein the further compound or composition comprises G-CSF.

18. Method according to any one of claims 1 to 3 wherein growth-hormone or one of its derivatives or any factor inducing growth hormone release is administered in an amount comprised between 20 to 50 μ g/kg of donor body weight, in an amount comprised between 30 to 40 μ g/kg of donor body weight or in an amount of 33 μ g per kilogram of donor body weight.

19. Method according to claim 17 wherein the G-CSF is administered in an amount comprised between 3 to 15 μ g/kg of donor body weight, in an amount comprised between 4 to 12 μ g/kg of donor body weight or in an amount of around 5 or 10 μ g per kilogram of donor body weight.

20. Method according to claim 2 wherein the administration of Growth Hormone is made three times a day and the administration of G-CSF is made daily.

21. Method according to any one of claims 1 to 3 wherein the administration of said composition is made by parenteral, subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal or buccal routes.

22. Method according to any one of claims 1 to 3 wherein the administration of said composition is daily or three times a day.

23. Method according to claim 1 or 2 wherein the administration of said composition is made over a period of 5 days or over a period of 10 days.

24. Method according to any one of claims 1 to 3 wherein the administration begins around 7 days after the beginning of a chemotherapeutic treatment or around 2 days after the end of a chemotherapeutic treatment.

25. Method according to any one of claims 1 to 3 wherein the growth hormone is recombinant growth hormone.

26. Method according to any one of claims 1 to 3 wherein the growth hormone is human growth hormone.

27. Method for the enhancement of hematopoiesis reconstitution in a human being comprising the steps of :

- a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo.

b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

c) transplanting the cells recovered in step (b) to an individual, and

d) administering to this individual growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to accelerate hematopoietic recovery.

28. Method according to claim 27 wherein administration in steps a) and d) further encompasses G-CSF.

29. Use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament which increases, in a donor, the number of circulating cells capable of regenerating hematopoiesis in vivo, said cells being used to treat by re-infusion, transplantation or engraftment a recipient in need of such cells.

30. Use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament to increase in a donor the number of circulating cells capable of regenerating hematopoiesis in vivo available for leukapheresis and re-infusion, transplantation or engraftment in a recipient in need of such cells.

31. Use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament, administered separately or simultaneously with G-CSF, to increase the number of circulating cells capable of regenerating hematopoiesis in vivo in a human being.

32. Use of human growth hormone or one of its derivatives or any factor inducing human growth hormone

release to prepare a medicament to increase over a period of up to 20 days the number of circulating cells capable of regenerating hematopoiesis in vivo in a human being.

33. Use according to any one of claims 29 to 32 wherein the medicament comprises one or several further compound(s) chosen among the following groups of compounds: hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

34. Use according to any one of claim 29, 30 or 32 wherein the medicament is administered separately or simultaneously with one or more further medicaments comprising one or several compound(s) chosen among the following groups of compounds: hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

35. Use according to claims 33 or 34 wherein the cytokines group comprises IL-1, IL-3, IL-6, IL-11, Insulin-like growth factor 1 (IGF-1), G-CSF, GM-CSF or SCF ; the chemokines group comprises MIP-1 α or thrombopoietin (TPO) ; the monoclonal antibodies group comprises anti-VLA-4 antibodies.

36. Use according to any one of claims 29 to 32 wherein the further compound or medicament comprises G-CSF.

37. Use according to any one of claims 29 to 32 wherein the administration of said medicament is made by parenteral, subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal or buccal routes.

38. Use according to any one of claims 29 to 32 wherein the administration of said medicament is daily or three times a day.

39. Use according to claim 31 wherein the administration of medicament comprising growth hormone

is made three times a day and the administration of G-CSF is daily.

40. Use according to any one of claims 29 to 32 wherein the administration of said medicament is made over a period of 3 days and/or until leukapheresis.

41. Use according to any one of claims 29 to 32 wherein the administration of said medicament begins around 7 days after the beginning of a chemotherapeutic treatment or around 2 days after the end of a chemotherapeutic treatment.

42. Use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing and/or treating opportunistic infections after transplantation or for limiting the risk of tumor recurrence after transplantation.

43. Use according to any one of claims 29 to 32 wherein growth hormone is recombinant growth hormone.

44. Use according to any one of claims 29 to 32 wherein growth hormone is human growth hormone.

45. Use according to any one of claims 29 to 32 wherein the circulating cells capable of regenerating hematopoiesis in vivo are CD34⁺ cells.

46. Use according to claim 45 wherein the increased number of CD34⁺ cells is more than 10, 25, 34 or 80 CD34⁺ cells per microliter of peripheral blood after administration of the medicament.

47. Use according to claim 45 wherein the increased number of CD34⁺ cells is at least 2×10^6 , 4×10^6 , 5×10^6 , 6×10^6 , 8×10^6 , 15×10^6 CD34⁺ cells per kilogram of body weight after administration of the medicament.

48. Use according to any one of claims 29 to 32 wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to around or more than 500 CFU-GM per milliliter of peripheral blood after administration of the medicament.

49. Use according to any one of claims 29 to 32 wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to an increased level of CFU-C, CFU-Meg or BFU-E in peripheral blood after administration of the medicament.

50. Use according to any one of claims 29 to 32 wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo substantially corresponds to a white blood cell count of around or more than 1000 cells per microliter of peripheral blood after administration of the medicament.

51. Use according to any one of claims 29 to 32 wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to around or more than 1×10^5 GM-CFC per kilogram of body weight.

52. Use according to any one of claims 29 to 32 wherein the circulating cells capable of regenerating hematopoiesis in vivo are CD34⁺/CD33⁺ cells and/or CD34⁺/CD38⁺ cells and/or CD34⁺/Thy-I cells and/or CD34⁺/Thy-I/CD38⁺ cells and/or CD33⁺ cells and/or bone-marrow stem cells and/or progenitor cells and/or long-term culture initiating cells (LTC-IC) and/or cells that fulfill self renewal potential and/or cells that fulfill pluripotential characteristics and/or cells that initiate long term bone marrow culture and/or cells that can generate multiple cell lineages.

53. Use according to any one of claims 29 to 32 wherein the target number of circulating cells capable of regenerating hematopoiesis in vivo is at least 2×10^4 LTC-IC per kg of donor or recipient body, around or more than 2×10^6 CD34⁺ cells per kilogram of donor or recipient body weight, around or more than 4×10^6 CD34⁺ cells per kilogram of donor or recipient body weight or around or more than 8×10^6 CD34⁺ cells per kilogram of body weight.

54. Use according to any one of claims 29 to 32 wherein growth-hormone or one of its derivatives or any factor inducing growth hormone release is administered in an amount comprised between 20 to 50 $\mu\text{g/kg}$ of body weight, in an amount comprised between 30 to 40 $\mu\text{g/kg}$ of body weight or in an amount of 33 μg per kilogram of body weight.

55. Use according to claim 36 wherein the G-CSF is administered in an amount comprised between 3 to 15 $\mu\text{g/kg}$ of body weight, in an amount comprised between 4 to 12 $\mu\text{g/kg}$ of body weight or in an amount of around 5 or 10 μg per kilogram of body weight.

1/2

Agents administered after chemotherapy		Absolute peak number of circulating CD34 ⁺ /μL				Total amount of leukapheresed CD34 ⁺ cells/kg of body weight			
G-CSF	Cycle	Patient	Patient	Patient	Patient	Patient	Patient	Patient	Patient
		1	2	3	10	12	50	80	25
GH + G-CSF	Cycle 1	10	12	50	25	34	80	5.10 ⁶	14.9 10 ⁶
	Cycle 2	25	34	80	5.10 ⁶	14.9 10 ⁶	6.1 10 ⁶	5.10 ⁶	6.1 10 ⁶

Figure 1

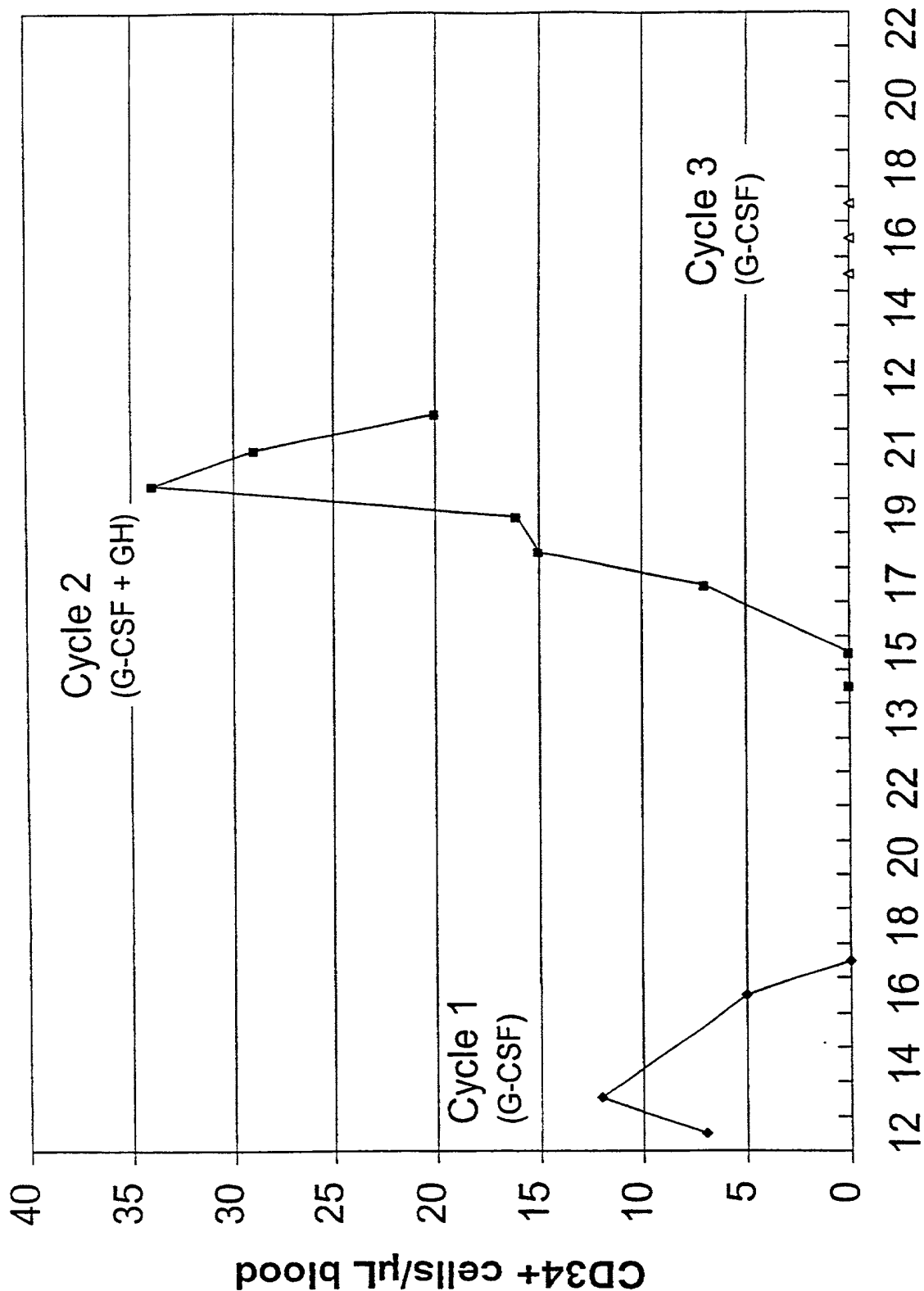


Figure 2
Days post-therapy

Combined Declaration for Patent Application and Power of Attorney

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name; and that I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

HUMAN GROWTH HORMONE TO STIMULATE MOBILIZATION OF PLURIPOTENT HEMATOPOIETIC STEM CELLS

the specification of which (check one)

- [] is attached hereto;
[] was filed in the United States under 35 U.S.C. §111 on _____, as U.S. Appln. No. _____*; or
[x] was/will be filed in the U.S. under 35 U.S.C. §371 by entry into the U.S. national stage of an international (PCT) application, PCT/EP99/10470; filed December 30, 1999, entry requested on _____*; national stage application received U.S. Appln. No. _____*; §371/§102(e) date _____* (* if known)

and was amended on _____ (if applicable).
(include dates of amendments under PCT Art. 19 and 34 if PCT)

I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above; and I acknowledge the duty to disclose to the Patent and Trademark Office (PTO) all information known by me to be material to patentability as defined in 37 C.F.R. §1.56.

I hereby claim foreign priority benefits under 35 U.S.C. §§ 119 (a)-(d) and 365 (b) of any prior foreign application(s) for patent, inventor's or plant breeder's rights certificate(s), or under §365(a) of any PCT application which designated at least one country other than the U.S., listed below:

Application No.	Country	Filing Date (MM/DD/YYYY)
98124834.7	EP	December 30 1998

If I claimed foreign priority above, I hereby identify below any foreign application for patent (including an international (PCT) application designating a country other than the United States) or for an inventor's or plant breeder's certificate, having a filing date before that of the earliest application from which foreign priority is claimed (if left blank, then there are none):

Non-Priority Application No	Country	Filing Date (MM/DD/YYYY)
_____	_____	_____
_____	_____	_____

I hereby claim the benefit under 35 U.S.C. §119(e) of any United States provisional applications listed below:

Application No.	Filing Date (MM/DD/YYYY)
_____	_____
_____	_____

I hereby claim the benefit under 35 U.S.C. §120 of any prior U.S. non-provisional application(s) or under §365(c) of any prior PCT international application(s) designating the U.S., listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in such U.S. or PCT international application in the manner provided by the first paragraph of 35 U.S.C. §112, I acknowledge the duty to disclose to the PTO all information which is material to patentability as defined in 37 C.F.R. §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

Application No	Filing Date (MM/DD/YYYY)	Status (patented, pending, abandoned)
_____	_____	_____
_____	_____	_____

As a named inventor, I hereby appoint the following registered practitioners to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

All of the practitioners associated with Customer Number 001444

Direct all correspondence to the address associated with Customer Number 001444, which is presently:

BROWDY AND NEIMARK, P.L.L.C.
624 Ninth Street, N.W.
Washington, D.C. 20001-5303
(202) 628-5197

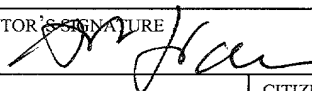
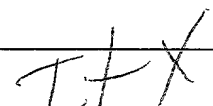
Title: _____

U.S. Application filed _____, Serial No. _____

PCT Application filed December 30, 1999, Serial No. PCT/EP99/10470

The undersigned hereby authorizes the U.S. Attorneys or Agents appointed herein to accept and follow instructions from SERONO INTERNATIONAL S.A. as to any action to be taken in the U.S. Patent and Trademark Office regarding this application without direct communication between the U.S. Attorneys or Agents and the undersigned. In the event of a change of the persons from whom instructions may be taken, the U.S. Attorneys or Agents appointed herein will be so notified by the undersigned.

I hereby further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C. §1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

FULL NAME OF FIRST INVENTOR <u>Alessandro Massimo GIANNI</u>	INVENTOR'S SIGNATURE 	DATE <u>Oct 10, 2001</u>
RESIDENCE <u>Italy</u>	CITIZENSHIP <u>Italy</u>	
POST OFFICE ADDRESS <u>Via Ludovico Muratori 29, 20135 Milano</u> 		
FULL NAME OF SECOND JOINT INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP	
POST OFFICE ADDRESS		
FULL NAME OF THIRD JOINT INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP	
POST OFFICE ADDRESS		
FULL NAME OF FOURTH JOINT INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP	
POST OFFICE ADDRESS		
FULL NAME OF FIFTH JOINT INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP	
POST OFFICE ADDRESS		
FULL NAME OF SIXTH JOINT INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP	
POST OFFICE ADDRESS		

ALL INVENTORS MUST REVIEW APPLICATION AND DECLARATION BEFORE SIGNING. ALL ALTERATIONS MUST BE INITIALED AND DATED BY ALL INVENTORS PRIOR TO EXECUTION. NO ALTERATIONS CAN BE MADE AFTER THE DECLARATION IS SIGNED. ALL PAGES OF DECLARATION MUST BE SEEN BY ALL INVENTORS.